

## REVIEW ARTICLE

# Protein kinase C pharmacology: refining the toolbox

Alyssa X. WU-ZHANG and Alexandra C. NEWTON<sup>1</sup>

Department of Pharmacology, University of California, San Diego, La Jolla, CA 92093-0721, U.S.A.

PKC (protein kinase C) has been in the limelight since the discovery three decades ago that it acts as a major receptor for the tumour-promoting phorbol esters. Phorbol esters, with their potent ability to activate two of the three classes of PKC isoenzymes, have remained the best pharmacological tool for directly modulating PKC activity. However, with the discovery of other phorbol ester-responsive proteins, the advent of various small-molecule and peptide modulators, and the need to distinguish isoenzyme-specific activity, the pharmacology of PKC has become increasingly complex. Not surprisingly, many of the compounds originally touted as direct modulators of PKC have subsequently been shown to hit many other cellular targets and, in some cases, not even directly modulate PKC. The complexities

and reversals in PKC pharmacology have led to widespread confusion about the current status of the pharmacological tools available to control PKC activity. In the present review, we aim to clarify the cacophony in the literature regarding the current state of *bona fide* and discredited cellular PKC modulators, including activators, small-molecule inhibitors and peptides, and also address the use of genetically encoded reporters and of PKC mutants to measure the effects of these drugs on the spatiotemporal dynamics of signalling by specific isoenzymes.

**Key words:** kinase activator, kinase inhibitor, kinase reporter, phorbol ester, protein kinase C (PKC).

## INTRODUCTION

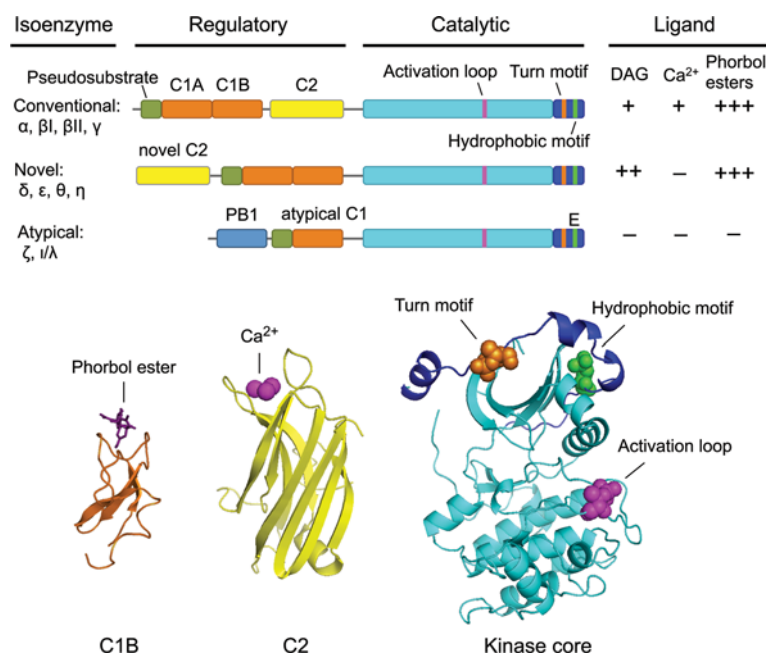
PKC (protein kinase C) isoenzymes transduce a wide range of extracellular signals that result in the generation of the lipid second messenger DAG (diacylglycerol), thereby regulating diverse cellular behaviours such as survival, growth and proliferation, migration, and apoptosis; consequently, their dysregulation is associated with a plethora of pathophysiologies. PKCs were famously discovered three decades ago to be direct signal transducers for a class of plant-derived tumour-promoting compounds called phorbol esters [1], which potently mimic the function of the endogenous ligand DAG [2]. Within the kinome, the PKC family belongs to the larger AGC family of kinases, named for protein kinases A, G, and C and also encompassing the related kinases protein kinase N, Akt, S6 kinase and PDK-1 (phosphoinositide-dependent kinase-1) [3].

The PKC family is composed of nine genes encoding ten well-characterized full-length mammalian isoenzymes that serve different biological roles, are regulated differently, and are classified as either conventional, novel or atypical according to the nature of their regulatory domains [3–9] (Figure 1). Conventional isoenzymes ( $\alpha$ , the alternatively spliced  $\beta$ I and  $\beta$ II, and  $\gamma$ ) each possess tandem C1A and C1B domains that bind to DAG or phorbol esters in membranes and a C2 domain that also binds membranes in the presence of the second messenger  $\text{Ca}^{2+}$ . Novel isoenzymes ( $\delta$ ,  $\epsilon$ ,  $\theta$  and  $\eta$ ) likewise each contain two tandem C1 domains that bind to DAG or phorbol esters, but possess a novel C2 domain that does not bind  $\text{Ca}^{2+}$  and does not serve as a membrane-targeting module; to compensate for the lack of

contribution of the C2 domain in membrane recruitment, the C1B domain of novel isoenzymes has a 100-fold higher affinity for DAG compared with the C1B domain of conventional PKCs [10,11]. Atypical isoenzymes ( $\zeta$  and  $\iota/\lambda$ ) do not respond to either DAG or  $\text{Ca}^{2+}$ ; rather, they possess a single atypical C1 domain that retains the ability to bind anionic phospholipids and a PB1 domain that mediates protein–protein interactions. Finally, the regulatory moiety of all these isoenzymes contains a short autoinhibitory pseudosubstrate sequence whose occupation of the kinase substrate-binding cavity maintains these kinases in an inactive state. Alternative transcripts beyond these ten isoenzymes exist, most notably the brain-specific PKM $\zeta$  (protein kinase M $\zeta$ ), which consists of the catalytic domain of PKC $\zeta$  [12] and PKC $\delta$  variants [13–15]. The catalytic moiety of all PKCs consists of a conserved kinase domain followed by a C-terminal tail. PKCs are constitutively processed by three ordered and tightly coupled phosphorylations in the catalytic domain that serve to mature the enzymes into a catalytically competent, but inactive and closed, conformation, in which the pseudosubstrate occupies the substrate-binding cavity. These phosphorylation sites are the activation loop, phosphorylated by the upstream kinase PDK-1, and two C-terminal sites termed the turn motif and hydrophobic motif. An exception exists in the case of atypical PKCs, which possess a phosphomimetic residue at the hydrophobic motif site. Canonically, PKCs are activated not by phosphorylation at these sites, which occurs constitutively, but by their acute translocation to membranes via second messenger-mediated membrane binding by their regulatory domains, an event that allosterically removes the pseudosubstrate from the active site. Three PKC isoenzymes

Abbreviations used: AKAP, A-kinase anchoring protein; CaMK,  $\text{Ca}^{2+}$ /calmodulin kinase; Cdc37, cell division cycle 37; CFP, cyan fluorescent protein; CHK, checkpoint kinase; CKAR, C-kinase activity reporter; DAG, diacylglycerol; FRET, fluorescence resonance energy transfer; GSK3, glycogen synthase kinase 3; Hsp90, heat-shock protein of 90 kDa; KSR1, kinase suppressor of Ras 1; LTP, long-term potentiation; MAPKAP-K, mitogen-activated protein kinase-activated protein kinase; mTOR, mammalian target of rapamycin; mTORC2, mTOR complex 2; PAK, p21-activated kinase; PDBu, phorbol 12,13-dibutyrate; PDK-1, phosphoinositide-dependent kinase-1; PHLPP, pleckstrin homology domain leucine-rich repeat protein phosphatase; PKA, protein kinase A; PKC, protein kinase C; PKD, protein kinase D; PKM $\zeta$ , protein kinase M $\zeta$ ; PP, pyrazolo[3,4-d]pyrimidine; RACK, receptor for activated C-kinase; RICK, receptor for inactive C-kinase; RSK, ribosomal S6 kinase; siRNA, small interfering RNA; YFP, yellow fluorescent protein; ZIP,  $\zeta$ -inhibitory peptide.

<sup>1</sup> To whom correspondence should be addressed (email anewton@ucsd.edu).



**Figure 1** Structure and relative ligand-responsiveness of PKC isoenzymes

Domain structure of PKC family members, showing autoinhibitory pseudosubstrate segment (olive green), C1 domains (orange), C2 domain (yellow), kinase core (cyan) and C-terminal tail (dark blue); the activation loop (pink), turn motif (orange) and hydrophobic motif (green) phosphorylation sites are also indicated. The ligands that bind to each subfamily, along with their relative affinities, are indicated. Also shown are the crystal structures of the C1B (PDB code 1PTR) [50], C2 (PDB code 1A25) [171] and kinase (PDB code 210E) domains [80].

(PKC $\alpha$ , PKC $\zeta$  and PKC $\iota/\lambda$ ) also possess C-terminal PDZ ligands that bind PDZ domain-containing protein scaffolds [16,17]. The structure, activation, life cycle, regulation and signalling dynamics of PKC isoenzymes have been reviewed abundantly elsewhere [3–9,18–21] and will not be addressed further in the present review.

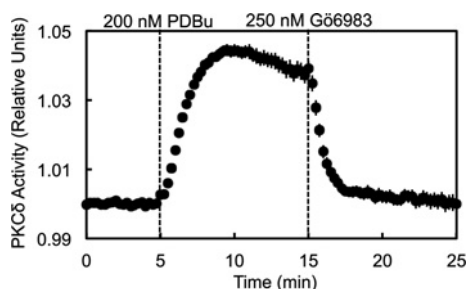
The pervasiveness of PKC as a signal transducer in so many biological contexts naturally causes its dysregulation to be implicated in a variety of diseases, such as cancer [4] and diabetes [22]. Thus elucidating the molecular mechanisms of PKC signalling has proven valuable to an understanding of both basic signalling biology and disease mechanism in a wide range of fields. However, one consequence of the extensive reach of research involving PKC as well as of the complexity inherent in studying a family of nine genes with diverse roles and modes of regulation has been a confusion about the current status of the pharmacological tools available to modulate PKC activity, especially with regard to which compounds are currently considered to be *bona fide* PKC modulators and which compounds have been discredited. The present review is our attempt to clarify the current status of the PKC pharmacological toolbox, including tools both for monitoring and for modulating PKC activity in cells.

## GENETICALLY ENCODED REPORTERS

The advent of genetically encoded FRET (fluorescence resonance energy transfer)-based reporters has enabled PKC pharmacology to be studied directly *in situ*. Although *in vitro* kinase activity assays have been the mainstay of PKC activity analysis for decades and continue to be the favoured method for *in vitro* profiling of PKC pharmacology, the cellular pharmacology of a kinase cannot always be reliably inferred from its *in vitro* profile [23]. Thus cellular assays should be performed to validate the effectiveness and specificity of a pharmacological reagent

against its intended target before its physiological effects are attributed to that target [24]. Although cellular PKC activity can be assayed biochemically by immunoblotting cell lysates with phosphorylation-specific antibodies against PKC substrates, genetically encoded reporters provide real-time spatiotemporal information on the effects of pharmacological tools on basal and agonist-evoked PKC activity in live cells.

CKAR (C-kinase activity reporter), the first and most commonly used PKC activity reporter [25,26], employs the modular architecture of activity reporters originally developed by Tsien and co-workers for tyrosine kinases [27] and for PKA (protein kinase A) [AKAR (A-kinase activity reporter)] [28], and subsequently also used in activity reporters for Akt/PKB (protein kinase B) [BKAR (B-kinase activity reporter)] [29] and PKD (protein kinase D) [DKAR (D-kinase activity reporter)] [30]. These reporters comprise a donor–acceptor FRET pair [typically CFP (cyan fluorescent protein) and YFP (yellow fluorescent protein)] flanking a kinase-specific substrate sequence tethered by a flexible linker to a phospho-peptide-binding module capable of binding the phosphorylated substrate sequence (reviewed in [31,32]). Phosphorylation of the substrate sequence causes the phospho-peptide-binding module to bind the substrate sequence, resulting in a conformational change in the reporter that produces a FRET change. In the case of CKAR, the unphosphorylated reporter experiences constitutive intramolecular FRET; however, upon phosphorylation of a threonine residue in the consensus substrate sequence by PKC, the substrate sequence binds to a phospho-threonine-binding FHA2 (forkhead-associated 2) domain, leading to conformational changes in the reporter that decreases its intramolecular FRET. This change in FRET is reversed upon dephosphorylation of the substrate peptide by cellular phosphatases. Thus changes in the FRET of CKAR serve as a readout for PKC activity in live cells in real time (Figure 2). CKAR is an effective reporter for the activity of all PKC isoenzymes [33], including atypical PKCs



**Figure 2** FRET ratio readout from a CKAR specific for PKC $\delta$  activity ( $\delta$ CKAR) reflects changes in live-cell PKC $\delta$  activity

COS-7 cells co-transfected with  $\delta$ CKAR and PKC $\delta$ -RFP (red fluorescent protein) were monitored for FRET ratio changes over time in response to stimulation with 200 nM of the phorbol ester PDBu and subsequent inhibition with 250 nM of the bisindolylmaleimide general PKC inhibitor Gö6983. FRET ratio traces recorded from 13 cells over three independent experiments were normalized to baseline data and plotted as the means  $\pm$  S.E.M. as a direct reflection of cellular PKC $\delta$  activity.

[23]. Modification of the substrate sequence has resulted in the generation of a PKC $\delta$ -specific reporter, which has been invaluable for identifying novel signalling mechanisms of this isoenzyme to the nucleus and mitochondria [33,34]. PKC activity can also be read out by measuring the phosphorylation-dependent release of the effector domain of MARCKS (myristoylated alanine-rich C-kinase substrate), an abundant PKC substrate that is released from the plasma membrane upon phosphorylation by PKC [35].

Because CKAR is a genetically encoded reporter, it can be positioned at specific locations within the cell through the use of appropriate targeting sequences or by fusion to specific proteins. Thus CKAR has been targeted to the plasma membrane, Golgi, nucleus, cytoplasm and mitochondria using short localization sequences [20,31,36]. It has also been targeted to protein scaffolds to monitor kinase activity on these context-specific signalling hubs [37,38]. This spatially resolved profiling of the rate, magnitude and duration of PKC activation has provided important insights into the subcellular contexts of PKC signalling, revealing spatiotemporal differences in PKC signalling in the presence of PKC inhibitors, PKC activators or phosphatase inhibitors.

FRET also provides a sensitive tool to monitor the translocation of PKC to specific cellular membranes, a hallmark of its activation. Specifically, intermolecular FRET reporters consisting of subcellularly targeted CFP and separate YFP-tagged PKC constructs have been used to monitor the translocation of PKCs to various intracellular organelles. This approach was used to unveil a novel isoenzyme-specific mechanism by which PKC $\delta$  interacts with mitochondria to promote mitochondrial respiration [34]. This approach has also been employed in a DAG reporter that uses the translocation of a YFP-tagged C1 domain to monitor the generation of DAG at various membranes, thereby revealing a rapid but transient rise in agonist-evoked DAG at the plasma membrane compared with a slower but sustained rise in DAG at the Golgi transduced by Ca<sup>2+</sup> [20,36,39]. A variety of other reporters for the PKC second messengers DAG and Ca<sup>2+</sup> are also available [26,40,41], including a recently developed reporter that simultaneously detects DAG and Ca<sup>2+</sup> [42]. The advantages of these FRET reporters over other techniques used to study molecular interactions and localization, such as pull downs, fractionations and immunofluorescence, include their preservation of the physiological cellular context and spatiotemporal information, their greater sensitivity for low affinity or transient interactions, and their accommodation of experimental adjustments in response to real-time data.

## ACTIVATORS

In contrast with the related Akt, which is acutely phosphorylated at the activation loop and hydrophobic motif upon agonist stimulation and whose activity can consequently be gauged using phosphorylation-specific antibodies against these sites, PKC is constitutively processed by phosphorylation into a catalytically competent but latent state [18]. Thus immunoblotting for PKC phosphorylation at its activation loop, turn motif or hydrophobic motif is not a reliable way to measure its activity. Rather, acute activation of PKC is achieved by the ligand-mediated engagement of its regulatory domains at membranes, which allosterically releases the autoinhibitory pseudosubstrate from its active site, shifting the enzyme into an active and open conformation. Thus the majority of compounds that directly activate PKC act as ligands for its regulatory domains and recruit PKC to cellular membranes.

In all except the atypical PKC isoenzymes, tandem C1 domains serve as hydrophobic switches [43] that drive PKC translocation to membranes. Specifically, each C1 domain contains a hydrophilic cleft in an otherwise hydrophobic surface. A variety of amphipathic ligands can insert their hydrophilic moieties into this hydrophilic cleft while their hydrophobic moieties complete a hydrophobic surface on the C1 domain with high affinity for membranes (Figure 1). Although the isolated C1A and C1B domains bind ligands, only one C1 domain is engaged on membranes in the context of full-length PKC; thus the stoichiometry of ligand binding is 1 mol of phorbol ester per mol of PKC [44–46]. This section will address the most prominent C1 domain ligands, the endogenous DAG and the phorbol esters, as well as a variety of other structurally diverse C1 domain ligands isolated from many different organisms. Structural comparison of these diverse ligands has led to a pharmacophore model for PKC ligands, identifying a set of functional groups at structurally homologous positions that is required for a ligand to interact with a PKC C1 domain [43]. Differences in the hydrophobic side chains projecting from these diverse ligands may give rise to different orientations of the C1 domain relative to the membrane, selectivity for different lipid microdomains and different biological responses as a consequence [43]. Although PKC may be the most prominent family of C1 domain-containing proteins, one should keep in mind that six other protein families representing 18 different signalling proteins, consisting of kinases, GTPase-regulating proteins and the signal-terminating DAG kinases, also respond to C1 ligands [47].

## Diacylglycerol

The canonical generation of DAG, the endogenous C1 ligand, occurs when receptor-stimulated PLC (phospholipase C) hydrolyses plasma membrane PtdIns(4,5)P<sub>2</sub> into the membrane-embedded DAG and the water-soluble Ins(1,4,5)P<sub>3</sub>. The consequent Ins(1,4,5)P<sub>3</sub>-induced increase in intracellular Ca<sup>2+</sup> subsequently activates phospholipases at distal membranes such as the Golgi, resulting in Ca<sup>2+</sup>-dependent elevation of DAG at the Golgi [39]. Thus investigators should be aware that pharmacological manipulations elevating intracellular Ca<sup>2+</sup> also elevate DAG. Although DAG has a simple structure, with only one chiral center, it can have many different combinations of fatty acids attached at the C-1 and C-2 positions. Short-chain synthetic DAGs such as DiC<sub>8</sub> (dioctanoyl) glycerol and OAG (oleoacetyl glycerol) are examples of more water-soluble versions of the generally very lipophilic DAG that effectively partition into cell membranes and can be used to transiently activate PKC. Synthetic

DAG-lactones with the structure of the DAG analogue constrained into the bound conformation have been designed for increased C1 binding [43]. DAG kinases, lipid kinases that phosphorylate DAG, converting it into PA (phosphatidic acid), terminate DAG signalling.

### Phorbol esters

Phorbol esters derive from the oil of the seed of the plant *Croton tiglium*, which has been used in traditional Chinese medicine, as a counter-irritant and cathartic in the 19th Century, and as a tumour promoter on the skin of mice in the mid-20th Century [43]. The active components in croton oil were purified in the late 1960s as 11 diesters of a tetracyclic diterpene called phorbol, with different combinations of fatty acids attached at the 12th and 13th numbered carbons [48]. Among these, PMA [phorbol 12-myristate 13-acetate, also known as TPA (12-*O*-tetradecanoylphorbol 13-acetate)] was found to be the most potent and became the paradigmatic tumour-promoting phorbol ester [43]. PDBu (phorbol 12,13-dibutyrate) was a derivative optimized for potency and reduced lipophilicity (and thus better aqueous solubility) that then enabled Driedger and Blumberg [49] to demonstrate the existence of a specific saturable receptor for phorbol esters, which led to the identification by Nishizuka and co-workers of PKC as the phorbol ester receptor [1]. This established the importance of PKC in cell proliferation and cancer and the employment of phorbol esters as a tool for activating PKC in cells.

As a tool for activating PKC, phorbol esters are much more potent than DAG: they bind PKC with two orders of magnitude higher affinity than DAG [44] and, additionally, are not readily metabolized (Figure 1). Thus low nanomolar concentrations of phorbol esters suffice to produce cellular actions [43]. Their remarkable potency is illustrated by the finding that 1 mol% PMA in the lipid bilayer increases PKC's membrane association by a staggering 4 orders of magnitude [44]. Similar to DAG, phorbol esters insert into the hydrophilic cleft of C1 domains, capping the C1 domain with a contiguous hydrophobic surface that retains the C1 domain on membranes [50,51].

Phorbol esters with different substituents on the 12th and 13th carbons can produce different biological responses, and not all of them promote tumorigenesis. For example, among 12-deoxyphorbol 13-monoesters, long-chain substituted 13-tetradecanoate is both inflammatory and tumour promoting [52], whereas short-chain substituted phorbol esters are also inflammatory, but not tumour promoting [53]. Blumberg and co-workers showed that ligand hydrophobicity determines the pattern and kinetics of PKC $\delta$  translocation, in turn dictating biological response [54]. Thus the more hydrophobic 12-deoxyphorbol 13-tetradecanoate, which causes PKC $\delta$  to translocate first to plasma membrane and then to internal membranes, is tumour promoting, whereas the more hydrophilic 12-deoxyphorbol 13-phenylacetate, which causes the immediate translocation of PKC $\delta$  to internal membranes, inhibits tumour promotion. The effect of lipophilicity on biological function raises caution in using phorbol esters to understand PKC signalling evoked by DAG. Furthermore, it suggests that therapeutic compounds designed to target the C1 domain should more closely resemble DAG than phorbol, an approach taken by Marquez and Blumberg [55].

### Fluorescent phorbol esters

Natural and synthetic fluorescent phorbol esters have been used to probe the spatiotemporal dynamics of phorbol ester behaviour and its interaction with PKCs. The synthetic fluorescent phorbol ester

dansyl-PMA has been shown to enter freely into cells, to behave like PMA in competing with PDBu for binding and in stimulating biological activity, and to localize to the endoplasmic reticulum, Golgi, mitochondrial and nuclear membranes [56,57]. In addition, the sapintoxins, naturally occurring and potent phorbol esters derived from *Sapium* plants that induce inflammation, but are poor tumour promoters, can act as FRET energy acceptors from the tryptophan residues of PKCs upon binding [57].

Using a series of homologous phorbol esters that varied in lipophilicity on the basis of the length of a methylene spacer used to attach a fluorescent BODIPY<sup>®</sup> (4,4-difluoro-4-bora-3a,4a-diaza-s-indacene) label, Braun et al. [58] monitored the spatiotemporal dynamics of ligand uptake in live cells. They found that the greater the lipophilicity of a ligand, the slower its rate of uptake, with ligands possessing PMA-like lipophilicity requiring 30–60 min to equilibrate, whereas more hydrophilic ligands penetrated more quickly. However, the equilibrium distributions of all of the ligands were similar, localizing primarily to intracellular membranes and failing to accumulate visibly within either the nucleus or plasma membrane. When they simultaneously monitored the distribution kinetics of these ligands together with those of fluorescently tagged PKC $\alpha$  or PKC $\delta$ , they found that the presence of overexpressed PKC modified and presumably trapped the distribution of the ligands. More lipophilic ligands now accumulated first in the plasma membrane before distributing to internal membranes, whereas more hydrophilic ligands penetrated to internal membranes more quickly. Additionally, there were isoenzyme-specific differences. PKC $\delta$  colocalized well with phorbol esters throughout the time course. In contrast, PKC $\alpha$  localized to the plasma membrane regardless of the phorbol ester distribution, suggesting that PKC $\alpha$  localization is dominated by other factors.

### Other C1 domain ligands

A variety of structurally diverse natural products bind with high affinity to the hydrophilic cleft of the C1 domain and activate PKC, reflecting the widespread physiological importance of PKC signal transduction. Ingenol and daphnane esters are tricyclic diterpenes structurally related to phorbol esters [43]. Ingenol mebutate, derived from the sap of *Euphorbia peplis*, a plant in the same family as the croton, is used as a topical treatment for the premalignant skin condition actinic keratosis [59]. Mezerein and daphnetoxin are daphnane esters [43] derived from *Daphne gnidium*, a poisonous Mediterranean evergreen shrub. Lyngbyatoxin and teleocidins are indole alkaloids, and aplysiatoxin is a polyacetate [43], all derived from bacteria. Iridals are triterpenoids derived from iridaceous plants [43,60].

Bryostatins are macrocyclic lactones [43] originally isolated from the marine bryozoan *Bugula neritina* [61]. Bryostatins are large complex molecules, as shown by the fact that bryostatin 1 has 11 chiral centers [43]. When bound to a C1 domain, the large protruding rings of bryostatin form a cap that contributes to its unique biology [43,62]. Although bryostatins induce some of the same effects as phorbol esters, their effects tend to be more transient, and they actually antagonize the effects of phorbol esters in numerous biological contexts, including that of tumour promotion [43]. Although the mechanism of this antagonism is unclear, bryostatin 1 has been in clinical trials, primarily as an anti-cancer agent, but also as a treatment for Alzheimer's disease [21,63,64]. Unlike phorbol esters, bryostatin 1 differentially down-regulates different PKC isoenzymes, with its down-regulation of PKC $\delta$  in particular exhibiting a biphasic dose-dependence [65,66]. Furthermore, whereas most phorbol

esters induce PKC $\delta$  to translocate first to the plasma membrane, then to internal membranes, bryostatin 1 induces direct translocation to internal membranes [67]. Although one study showed that tethering PKC $\delta$  to the plasma membrane promoted apoptosis, mimicking the effects of phorbol esters [68], analysis of the effects of phorbol esters, whose lipophilicity covered eight orders of magnitude, revealed that parameters beyond lipophilicity and translocation pattern drive the biological differences between phorbol esters and bryostatins [69]. As bryostatins are difficult both to isolate from their non-renewable natural sources and to synthesize, investigators have designed simplified bryologues [70–72].

### Ca<sup>2+</sup> agonists

Elevation of intracellular Ca<sup>2+</sup> using either a natural agonist such as histamine or a pharmacological agent such as thapsigargin (an inhibitor of the sarco/endoplasmic reticulum Ca<sup>2+</sup>-ATPase) or ionomycin (a Ca<sup>2+</sup> ionophore) directly activates the Ca<sup>2+</sup>-sensitive conventional PKC isoenzymes by the binding of Ca<sup>2+</sup> to conventional C2 domains to target the domain to the PtdIns(4,5)P<sub>2</sub>-enriched plasma membrane via electrostatic interactions. Furthermore, as noted above, Ca<sup>2+</sup> stimulates phospholipase activity to produce DAG at internal membranes such as the Golgi [39], which activates both conventional and novel PKCs. Thus Ca<sup>2+</sup> elevation directly activates conventional PKC isoenzymes by recruiting them via their C2 domains to membranes and indirectly activates both conventional and novel PKCs by elevating DAG to recruit them via their C1 domains to membranes.

### Natural agonists

A variety of diverse physiological stimuli result in DAG production and consequently PKC activation. G-protein-coupled receptor agonists such as histamine, UTP, bombesin and lysophosphatidic acid, and receptor tyrosine kinase agonists such as insulin, epidermal growth factor, platelet-derived growth factor, fibroblast growth factor and other growth factors are commonly used to activate cellular PKC. The choice of agonist depends on the cell type and its receptor/signalling pathway signature. For example, in standard tissue-culture experiments, ATP activates PKC in MDCK (Madin–Darby canine kidney) cells [26], UTP activates PKC in COS-7 cells [33,36] and histamine activates PKC in HeLa cells [26].

## SMALL-MOLECULE INHIBITORS

### Active-site inhibitors

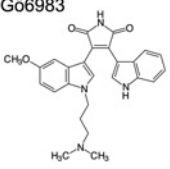
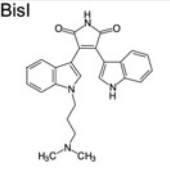
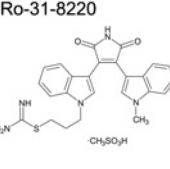
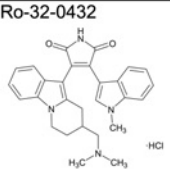
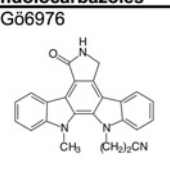
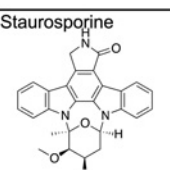
The vast majority of small-molecule kinase inhibitors target the ATP-binding site (Figure 3 and Table 1). When selecting from among the numerous such compounds marketed as PKC inhibitors, one should be aware of the selectivity of the inhibitor for PKCs over other kinases, the selectivity of the different PKC isoenzymes in general for being targeted by kinase inhibitors, and the meaning of the IC<sub>50</sub> value for that compound (Figure 3). First, all PKC inhibitors possess some degree of promiscuity [73–76]. Secondly, among the PKCs, the atypical isoenzymes are the most selective, with only 1–2% of compounds in a screen of 178 kinase inhibitors used at 500 nM inhibiting either PKC $\zeta$  or PKC $\iota$  [76] and less than 3% of compounds in a different screen of 72 kinase inhibitors used at 300 nM binding to PKC $\iota$  [75]. Conventional and novel PKCs have somewhat lower levels of

selectivity, with approximately 5–10% of compounds in either screen hitting these isoenzymes [75,76]. Thirdly, the reported IC<sub>50</sub> values of ATP-competitive inhibitors depend on the ATP concentration used in the assay [73]. This is an oft-neglected point: the intracellular concentration of ATP is in the order of 5 mM, the K<sub>m</sub> value for ATP for PKC isoenzymes is in the order of 5  $\mu$ M (see [77]) and the K<sub>i</sub> value for inhibitors is typically in the order of 5 nM (e.g. [78]). With these parameters, 1  $\mu$ M inhibitor may result in a 95% inhibition of kinase activity in an *in vitro* assay with 50  $\mu$ M ATP, but only 17% inhibition of activity in the presence of 5 mM ATP. The majority of effective PKC inhibitors in prevalent laboratory use today are ATP-competitive derivatives of bisindolylmaleimides or indolocarbazoles.

### Bisindolylmaleimides

Among the bisindolylmaleimides are Gö6983 and BisI (also known as Gö6850 or GF109203X), both ATP-competitive, isoenzyme-non-specific general PKC inhibitors that differ from one another by only a methoxy group (Figure 3). Both compounds inhibit PKC isoenzymes from all three classes *in vitro*, but are more effective against conventional and novel PKCs than atypical isoenzymes [76,78,79], exhibit a moderate degree of kinase promiscuity [76], and do not inhibit the closely related PKA or PKD [76,79]. Gö6983 inhibits PKC isoenzymes with IC<sub>50</sub> values ranging from 7 nM to 60 nM (at 35  $\mu$ M ATP) [79], whereas BisI inhibits PKC isoenzymes with IC<sub>50</sub> values ranging from the nanomolar to the micromolar (at 10  $\mu$ M ATP) [78]. At 500 nM, both Gö6983 and BisI inhibit conventional and novel PKCs by 80–100%, with Gö6983 being most effective against PKC $\delta$  and BisI being most effective against PKC $\epsilon$ , and both inhibit atypical PKCs by less than 50% [76]. In addition to inhibiting PKCs, both Gö6983 and BisI exhibit a moderate degree of promiscuity. At 500 nM, Gö6983 substantially inhibited (by >50%, greater than its inhibition of atypical PKCs) 21 other kinases in a panel of 300 mostly full-length kinases screened, whereas BisI substantially inhibited 27 [76]. Of these off-target hits, Gö6983 dramatically inhibited (by >84%, equal to or greater than its inhibition of conventional and novel PKCs) five of these kinases [GSK3 (glycogen synthase kinase 3)  $\alpha$  and RSK (ribosomal S6 kinase) 1–RSK4], whereas BisI dramatically inhibited ten [FLT3 (Fms-like tyrosine kinase-3), GSK3 $\alpha$ , GSK3 $\beta$ , S6 kinase, PIM1, PIM3 and RSK1–RSK4] [76]. At 1  $\mu$ M, BisI also strongly inhibited MAPKAP-K (mitogen-activated protein kinase-activated protein kinase) 1 $\beta$ , but was otherwise highly selective for PKC when tested against a panel of 24 kinases [73]. X-ray crystallography shows that BisI localizes to the ATP-binding pocket of the kinase domain of PKC $\beta$ II, where its dimethylamino group hydrogen bonds with the catalytic Asp<sup>470</sup> of PKC $\beta$ II [38,80].

A series of ATP-competitive bisindolylmaleimides originally synthesized by Roche are commercially marketed as PKC inhibitors [81]. Among these, Ro-31-8220 and Ro-32-0432 have been characterized in screens of kinase inhibitor selectivity (Figure 3). Both were originally reported to inhibit rat brain PKC with an IC<sub>50</sub> value of approximately 20 nM (at 10  $\mu$ M ATP) and each of the conventional PKC isoenzymes with an IC<sub>50</sub> value of between 5 and 37 nM, both with the highest preference for PKC $\alpha$  [81]. The major reported distinction between them was that Ro-32-0432 had a much higher IC<sub>50</sub> value for PKC $\epsilon$  (108 nM) than Ro-31-8220 did (24 nM) or than either had for conventional isoenzymes [81]. The specificity of Ro-31-8220 was screened at 100 nM, at which concentration it inhibited PKC $\alpha$  by 97% and PKC $\zeta$  by 75%, substantially inhibited (by

Inhibitor	IC <sub>50</sub>	Conc.	% Inhibition of PKC Isoenzymes			% Inhibition of Non-PKC Kinases (total tested)	
			Conventional	Novel	Atypical	Inhibited by ≥50%	Inhibited by ≥80%*
<b>Bisindolylmaleimides</b>							
 G66983	6-7 nM for α, β, γ, 10 nM for δ, 60 nM for ζ (35 μM ATP) [79]	500 nM [76]	80-100%		<50%	7.2% (290)	1.7% (290)
 BisI	8.4 nM for α, 18 nM for β, 210 nM for δ, 132 nM for ε, 5.8 μM for ζ (10 μM ATP) [78]	500 nM [76]	80-100%		<50%	9.3% (290)	3.4% (290)
		1 μM [73]	α by 96%	N.T.	N.T.	17% (23)	4.3% (23)
 Ro-31-8220	5-27 nM for cPKC, 24 nM for ε (10 μM ATP) [81]; 33 nM for PKCα (100 μM ATP) [73]	100 nM [82]	α by 97%	N.T.	ζ by 75%	22% (67)	13% (67)
		1 μM [73]	α by 97%	N.T.	N.T.	30% (23)	17% (23)
 Ro-32-0432	9-37 nM for cPKC, 108 nM for PKCε (10 μM ATP) [81]	500 nM [76]	>87%		64-69%	4.5% (290)	2.4% (290)
<b>Indolocarbazoles</b>							
 G66976	2.3 nM for α, 6.2 for β, 7.9 for rat brain PKC (10 μM ATP) [78]	100 nM [82]	α by 98%	N.T.	ζ by 22%	57% (67)	28% (67)
		500 nM [76]	α, β, βII by >80%, γ by 28%	δ, ε, η by 41-65%, θ by 24%	ζ by 13%, ι by 8%	37% (290)	18% (290)
		1 μM [73]	α by 93%	N.T.	N.T.	48% (23)	30% (23)
 Staurosporine	11-32 nM for cPKC, PKCε (10 μM ATP) [81]; 2.7 nM for rat brain PKC (5 μM ATP) [93]; not detected for PKCζ [78]	500 nM [76]	94-100%		ζ by 78%, ι by 95%	82% (290)	69% (290)

N.T. = not tested

\*Comparable to inhibition of PKCs

**Figure 3** Small-molecule active-site inhibitors: *in vitro* potency, isoenzyme selectivity and promiscuity

≥50%) 15 other kinases in a panel of 69 kinases screened, and dramatically inhibited (by ≥80%) nine of those [82]. At 1 μM, it inhibited PKCα by 97%, was found to have an IC<sub>50</sub> value of 33 nM for PKCα (at 100 μM ATP), substantially inhibited seven other kinases in a panel of 24 kinases screened and dramatically inhibited four of those [73]. These data are consistent with another report of the non-specificity of Ro-31-8220 that, similar to the aforementioned screening studies, also found MAPKAP-K1β and S6 kinase to be targets of this compound [83]. The specificity of Ro-32-0432 was screened at 500 nM, at which concentration it was found to be the 14th most selective inhibitor among 178 kinase inhibitors screened and the most selective PKC inhibitor of the seven included [76]. At 500 nM, Ro-32-0432 inhibited all conventional and novel PKC isoenzymes by >87% without exhibiting much isoenzyme specificity, inhibited atypical PKC

isoenzymes by 64–69%, substantially inhibited (by >50%) only 13 other kinases in a panel of 300 kinases screened, and dramatically inhibited (by >80%) only seven of those [76]. Incidentally, Ro-32-0432 is one of only two kinase inhibitors among 178 screened found to be able to inhibit PKCζ substantially [76].

A variety of other indolylmaleimide-based PKC inhibitors have been developed, including several that have been advanced for clinical development. Sotrastaurin (AEB071) is a novel indolylmaleimide that potently and selectively inhibits novel and conventional PKCs, with nanomolar or sub-nanomolar IC<sub>50</sub> values for these PKCs compared with above-micromolar or near-micromolar IC<sub>50</sub> values for atypical PKCs and all 32 other kinases screened (at 10 μM ATP) [84,85]. As an immunosuppressive that markedly inhibits PKCθ in T-cells and

**Table 1** Mechanism, efficacy *in vitro* compared with in cells and specificity of various inhibitor types

Inhibitor	Mechanism of action	Inhibition of PKC		Specificity for PKC
		<i>In vitro</i>	In cells	
Small molecule				
Active site	ATP-competitive	Yes	Yes	Variable, see Figure 3
Non-active site				
BisIV	Allosteric, uncompetitive with substrate	Yes	Yes	At 500 nM, inhibited 4.5 % of 290 other kinases by $\geq 50\%$ (as much as it inhibited PKCs) [76]
Calphostin C	Competes against DAG/phorbol ester binding to C1 domain	Yes	Yes	Not well characterized
PS168, PS171 benzimidazole compounds	Allosterically inhibits by binding PIF pocket	Yes	Yes	Selectively inhibits PKC $\zeta$ over other isoenzymes and AGC kinases [114]
Geldanamycin, 17-AAG, Celestrol	Inhibits PKC maturation by inhibiting Hsp90/Cdc37	No	Yes (not acute)	Not applicable
Torin1	Inhibits PKC maturation by inhibiting mTORC2	No	Yes (not acute)	Not applicable
Peptides				
ZIP and other pseudosubstrate-derived peptides	Highly basic peptide competes with substrate	Yes	No	None
Discredited				
Rottlerin	Mitochondrial uncoupler	No	No	None
Chelerythrine	Induces apoptosis	Yes/No	No	None

T-cell activation and proliferation [84], sotrastaurin has been in clinical trials for psoriasis [21,86] and transplant rejection [85]. Enzastaurin (LY317615) and ruboxistaurin (LY333531) are PKC $\beta$ -selective bisindolylmaleimides with IC<sub>50</sub> values of 4.7–6 nM for PKC $\beta$  (at 30  $\mu$ M ATP), 10- to >100-fold that for other PKC isoenzymes [87,88]. Both exhibit modest promiscuity. At 1  $\mu$ M, enzastaurin did not substantially inhibit any of the 28 other kinases tested [89] and ruboxistaurin is selective for PKC $\beta$  over PKA, CaMK (Ca<sup>2+</sup>/calmodulin kinase), Src and casein kinase [87] and, at 100 nM, substantially inhibited 12 other kinases in a panel of 69 kinases screened, dramatically inhibiting four of them [82]. Enzastaurin, which inhibits proliferation and induces apoptosis in human tumour cell cultures and xenografts, is in oncology clinical trials as an anti-angiogenic [21,82,89,90]. Ruboxistaurin was in clinical trials for the treatment of diabetic vascular dysfunction that were discontinued during Phase III [21,82,91]. Anilino-monoindolylmaleimides, compounds in which one of the indoles of bisindolylmaleimide has been replaced with aniline, are selective for PKC $\beta$  over PKC $\alpha$  or PKC $\epsilon$  by approximately one order of magnitude [88]. However, one such compound, termed simply 'PKC $\beta$  inhibitor', when used at 500 nM, was reported to also substantially inhibit PKC $\alpha$ , PKC $\eta$  and PKC $\theta$ , and to dramatically inhibit PKC $\delta$  and PKC $\epsilon$ ; nonetheless, Anastassiadis et al. [76] ranked this compound as the second most selective PKC inhibitor of the seven included, after Ro-32-0432.

#### Indolocarbazoles

Among the indolocarbazoles are the ATP-competitive inhibitors Gö6976 and staurosporine (Figure 3). Gö6976 inhibits conventional PKC isoenzymes at nanomolar concentrations *in vitro*, but does not effectively inhibit novel or atypical PKC isoenzymes even at micromolar concentrations [78]. The potency and specificity of Gö6976 has been characterized in multiple studies of kinase inhibitor selectivity [73,76,82]. At 100 nM, Gö6976 inhibited PKC $\alpha$  by 98 % and PKC $\zeta$  by only 22 %, and the other PKC isoenzymes were not tested [82]. At 500 nM, it inhibited PKC $\alpha$ , PKC $\beta$ I and PKC $\beta$ II by more than 80 %; PKC $\delta$ ,

PKC $\epsilon$  and PKC $\eta$  by 41–65 %; PKC $\gamma$  and PKC $\theta$  by less than 28 %; and atypical PKC isoenzymes by less than 13 % [76]. At 1  $\mu$ M, it inhibited PKC $\alpha$  by 93 %, and the other isoenzymes were not tested [73]. However, these studies also reveal that Gö6976 is a highly promiscuous inhibitor of other kinases. At 100 nM, it substantially inhibited (by  $\geq 50\%$ ) 38 other kinases in a panel of 69 kinases screened and dramatically inhibited (by >80 %) 19 of those [82]. At 500 nM, it was the seventh most promiscuous compound in a panel of 178 kinase inhibitors screened, substantially inhibiting (by >50 %) 107 kinases other than PKCs in a panel of 300 kinases screened and dramatically inhibiting (by >80 %) 52 of those [76]. At 1  $\mu$ M, Gö6976 substantially inhibited (by  $\geq 50\%$ ) 11 other kinases in a panel of 24 kinases screened and dramatically inhibited (by >80 %) seven of those [73]. The dramatically inhibited off-target hits of Gö6976 found by both the Cohen and Peterson groups are Aurora B, CaMKK2 (CaMK kinase 2), CDK2 (cyclin-dependent kinase 2)-cyclin A, CHK (checkpoint kinase) 1, GSK3 $\beta$ , MSK1 (mitogen- and stress-activated kinase 1), MST2, PAK (p21-activated kinase) 4, PAK5, PAK6, PHK, PIM3, PKD1 [79], RSK1 and RSK2 [73,76,82]. Parallel usage of Gö6976 and a general PKC inhibitor such as Gö6983 in an experiment can be employed to analyse whether a biological effect is the result of activity by an off-target kinase (inhibited by Gö6976, but not by Gö6983) or whether it is the result of novel or possibly atypical PKC activity (inhibited by Gö6983, but not by Gö6976).

Staurosporine represents the original indolocarbazole isolated from *Streptomyces staurosporeus* [92,93] and is a highly promiscuous, ATP-competitive kinase inhibitor [38,74–76] that potently binds to and inhibits most if not all PKC isoenzymes *in vitro* [74–76,81]. Indeed, staurosporine has been ranked as the most non-selective commercially available kinase inhibitor in multiple unbiased high-throughput screens of kinase inhibitor selectivity [74–76]. Despite its promiscuity and one report that staurosporine does directly inhibit PKC $\zeta$  *in vitro* [76], other *in vitro* studies report that staurosporine does not inhibit either PKC $\zeta$  [94] or PKM $\zeta$  [95], a brain-specific alternative transcript of PKC $\zeta$  encoding its catalytic domain [12]. Consequently, staurosporine has been used as a negative control inhibitor in

studies that implicate PKM $\zeta$  in long-term memory [95,96]. However, staurosporine does bind with nanomolar affinity to and inhibits PDK-1 [74,76], the upstream kinase that catalyses the activation loop phosphorylation required for the catalytic competence of all PKC isoenzymes [18,97], including that of PKC $\zeta$  [98,99]. Thus, even though staurosporine may not have inhibited PKM $\zeta$  *in vitro* in some studies, it does inhibit PKM $\zeta$  activity in cells and tissues, as measured both by using CKAR and by immunoblotting for the phosphorylation of endogenous PKM $\zeta$  substrates, by inhibiting PDK-1's phosphorylation of PKM $\zeta$ 's activation loop [23]. Effects such as this, which would be overlooked in *in vitro* assays, demonstrate the importance of verifying the targets of a drug in the complex milieu of cells and tissues before drawing physiological conclusions based solely on its *in vitro* pharmacological profile. Additionally, UCN01 (hydroxystaurosporine) is a staurosporine derivative that also potentially inhibits many kinases in addition to PKCs [73,82], does not inhibit PKC $\zeta$  *in vitro* [82,94], but does inhibit PDK-1 [73,82,100].

#### Active-site inhibitors with additional binding determinants

A recent chemical library screen identified thieno[2,3-D]pyrimidine-based compounds that bind competitively with respect to ATP in the nucleotide-binding pocket of atypical PKC isoenzymes and, additionally, have a benzyl group that displaces a key phenylalanine residue that structures the adenosine-binding motif [101]. Thus, not only do the compounds inhibit ATP binding, they also cause the adenosine-binding motif to become disordered. These promising compounds are effective in cells and serve as a paradigm for isoenzyme-selective inhibition by coupling active-site inhibition with isoenzyme-selective binding determinants.

#### Active site occupation by an inhibitor prevents PKC dephosphorylation

An interesting and counterintuitive effect of PKC active-site inhibitors is their stabilization of the constitutive processing phosphorylations on PKC [102,103]. Cameron et al. [102] first observed an increase in the steady-state levels of these processing phosphorylations on the normally unprimed kinase-dead constructs of PKC $\epsilon$  and PKC $\alpha$  upon incubation with ATP-competitive inhibitors such as BisI, Gö6983 or Gö6976, but not with the non-active-site inhibitor calphostin C. The same effect was observed by Okuzumi et al. [104] for the analogous regulatory phosphorylations on Akt. These observations were recapitulated for PKC $\beta$ II *in vitro* and in cells by Gould et al. [103], who further showed that the inhibitor-induced increase in steady-state levels of processing phosphorylations results from protection against dephosphorylation rather than promotion of phosphorylation. Specifically, pulse-chase analysis revealed that incubation with BisI had no effect on the rate of processing phosphorylations, but significantly slowed phorbol ester-triggered dephosphorylation. Similarly, occupation of the active site of Akt by ATP-competitive inhibitors or by ATP itself protects Akt from dephosphorylation by restricting phosphatase access [105,106]. Finally, the same protection from dephosphorylation can be observed upon active-site occupation by peptide substrates or by the pseudosubstrate peptide of PKC itself when in its inactive conformation [107,108]. At the structural level, active-site occupancy causes the C-terminal tail of PKC [80] and PKA [109,110], where the turn and hydrophobic motifs reside, to go from a highly flexible to a highly ordered state.

Scaffolded PKC is refractory to inhibition by active-site inhibitors

Protein scaffolds such as AKAPs (A-kinase anchoring proteins) and RACKs (receptors for activated C-kinase) organize PKCs, their substrates and their modulators into subcellular complexes that co-ordinate signalling. It was recently discovered that PKC bound to protein scaffolds such as AKAP79 is refractory to inhibition by ATP-competitive inhibitors such as BisI or staurosporine [38]. The probable explanation is that the apparent concentration of substrate, when anchored in such close proximity to an enzyme, is so high in the neighbourhood of the enzyme that active-site inhibitors become ineffective. Thus the context of PKC's interactions with protein scaffolds not only determines the spatiotemporal dynamics and substrate specificity of PKC in cells, but also appears to blunt the effectiveness of active-site inhibitors against PKC, shifting its pharmacological profile. In contrast, non-active-site inhibitors such as BisIV or calphostin C maintain their effectiveness against AKAP79-scaffolded PKC [38].

#### Non-active-site inhibitors

Although most drug development efforts against kinases have focused on the ATP-binding site, the kinome-wide conservation of the active site results in widespread drug promiscuity among active-site inhibitors. Structure-based design that targets less conserved pockets within the kinase domain or targeting non-catalytic domains to generate drugs that inhibit kinases by an allosteric mechanism could produce more selective kinase inhibitors. One example of a non-active-site PKC inhibitor is BisIV (Table 1). Similar to the active-site PKC inhibitor BisI, BisIV is a bisindolylmaleimide-derived general PKC inhibitor. But, unlike BisI, from which it differs only by the absence of the dimethylamino group that hydrogen bonds with the catalytic aspartate residue, BisIV is uncompetitive with respect to substrate [38]. Although BisIV is a much less potent inhibitor than BisI, Gö6983 or Gö6976, it is also more specific to PKC [73,76]. Because it is not an active-site inhibitor, despite its structural similarity to BisI, BisIV treatment does not stabilize PKC's priming phosphorylations [103] and remains effective against scaffolded PKC [38]. The same is true of calphostin C (Table 1), which targets the DAG-binding sites of PKC in a light-dependent manner [38,102,111,112]. Another example of allosteric PKC inhibition involves modulation of a site called the PIF-pocket in the N-lobe of AGC kinases, a site that binds the phosphorylated hydrophobic motif segment [113]. Biondi and co-workers have identified benzimidazole compounds (namely PS171 and PS168; Table 1) that allosterically inhibit PKC $\zeta$  *in vitro* by binding its PIF pocket and, additionally, inhibit PKC $\zeta$ -dependent NF- $\kappa$ B (nuclear factor  $\kappa$ B) activation in cells [113,114]. These allosteric modulators are the first *bona fide* inhibitors of PKC $\zeta$  shown to have cellular effects on functions mediated by this kinase.

#### Inhibitors of PKC maturation

Upon translation, conventional and novel PKCs are constitutively processed by phosphorylation at the activation loop, turn motif and hydrophobic motif to mature them into a stable and catalytically competent, but inactive, conformation [3,19]. Species that are not processed by phosphorylation are degraded by the proteasome. Thus the steady-state levels of conventional and novel PKC isoenzymes are extremely low in cells lacking the upstream kinase PDK-1 [115]. However, active-site inhibitors of PDK-1 are relatively ineffective at inhibiting the processing of these PKC isoenzymes, underscoring the concept that kinases scaffolded on/next to their substrates are refractory to active-site inhibitors [38]. In contrast, atypical PKC isoenzymes, which have a



phosphomimetic residue at the hydrophobic motif, are sensitive to inhibition by PDK-1, and phosphorylation of their activation loop acutely controls activity.

The initiation of PKC maturation is promoted by the chaperone Hsp90 (heat-shock protein of 90 kDa) and its co-chaperone Cdc37 (cell-division cycle 37), which bind to newly synthesized PKC via a molecular clamp formed between a conserved PXXP motif in the C-terminal tail of PKC and a conserved tyrosine residue in the  $\alpha$ E-helix of the kinase core [116]. Thus Hsp90 inhibitors, such as geldanamycin and 17-AAG, which inhibit the ATP binding and hydrolysis essential for Hsp90's protein folding function, and celastrol, which inhibits both Hsp90's ATP binding and its interaction with Cdc37, inhibit PKC hydrophobic motif phosphorylation and enzyme maturation [116] (Table 1).

Phosphorylations of both the turn and hydrophobic motifs have been shown to depend on mTORC2 [mTOR (mammalian target of rapamycin) complex 2] and do not occur in mTORC2-deficient cells [91,117–119], although it is unclear whether mTORC2 directly phosphorylates PKC at one or both of these sites or whether it performs a chaperoning role as Hsp90 does to prime PKC for phosphorylation by another mechanism, such as autophosphorylation, as observed *in vitro* [3]. Thus second-generation ATP-competitive mTOR inhibitors such as Torin1 can reduce the steady-state levels of PKC by preventing priming phosphorylations [120]; they do not, however, acutely inhibit PKC activity (Table 1).

#### Inhibitors of PKC down-regulation

In the mature but inactive conformation of PKC, its pseudosubstrate peptide is lodged in the kinase active site, preventing it from phosphorylating substrates. Agonist stimulation induces PKC to translocate to intracellular membranes, where membrane engagement of its regulatory domains allosterically removes the pseudosubstrate from the active site, enabling it to phosphorylate substrates. In this open conformation, PKCs become more sensitive to dephosphorylation by two orders of magnitude [107], and sustained activation of PKC causes it to become dephosphorylated and degraded, a process termed down-regulation [3,19]. The peptidyl-prolyl isomerase Pin1 controls the down-regulation of conventional PKCs by docking on to its hydrophobic motif and isomerizing the phospho-threonine–proline peptide bond of the turn motif from a *cis* to a *trans* conformation, converting PKC into a down-regulation-sensitive species [121]. Thus Pin1 inhibitors such as PiB (diethyl-1,3,6,8-tetrahydro-1,3,6,8-tetraoxobenzo[*lmm*])[3,8]phenanthroline-2,7-diacetate) inhibit the down-regulation of conventional PKCs and trap them in the active state [121]. PHLPP (pleckstrin homology domain leucine-rich repeat protein phosphatase) dephosphorylates the hydrophobic motif of conventional and novel PKCs, the first event in the down-regulation of PKC, whereas an okadaic acid-sensitive phosphatase dephosphorylates the turn motif and activation loop [122]. Thus phosphatase inhibitors that can inhibit PHLPP or the turn motif phosphatase or mutation of the hydrophobic motif to a phosphomimetic glutamate also inhibit down-regulation and trap PKC in the active state [122].

## PEPTIDE DRUGS

### Peptides derived from RACKs

PKC isoenzymes with overlapping activator responsiveness and substrate specificities frequently coexist in the same tissues, but

their distinct subcellular localizations account in part for their isoenzyme-specific functions [123]. The subcellular localizations of PKC isoenzymes are determined not only by lipid interactions, but also in part by their interactions with scaffold proteins such as RACKs, structurally unrelated adaptor or scaffold proteins that are functionally defined by their ability to bind and activate specific PKC isoenzymes and were first identified by Mochly-Rosen et al. [124] in the particulate fraction of cells over two decades ago. RACKs are known to bind the regulatory domains and variable N- and C-terminal sequences of PKCs and to activate them by competing for binding against autoinhibitory sequences within PKCs, relieving their autoinhibition and locking them into an active conformation that exposes their substrate-binding site [123,125]. For example, the first and most archetypal autoinhibitory sequence to be discovered in PKC is its pseudosubstrate peptide [126]. RACKs can thus control both PKC isoenzyme-specific localization (and hence substrate proximity) and activation. Another set of anchoring proteins was proposed to exist for inactive PKC isoenzymes [RICKs (receptors for inactive C-kinase)] [123].

Taking advantage of the isoenzyme-specific interactions between PKCs and RACKs to address the dearth of isoenzyme-specific PKC activators and inhibitors, Mochly-Rosen and co-workers developed a series of first-generation peptides, derived from either PKCs or RACKs and based on their interaction sites, that interfere with the protein–protein interactions between PKCs and RACKs [123,127]. In cases where a RACK acts to scaffold PKC next to its substrates, peptides that interfere with PKC binding to RACK will inhibit PKC signal transduction, and such peptides act as isoenzyme-specific inhibitors of PKC translocation and activity [123,127]. By the same principle, peptides that interfere with PKC–RICK interaction should act as isoenzyme-specific PKC agonists [123]. However, peptides called pseudo-RACKs ( $\psi$ RACKs) that mimic the autoinhibition-relieving properties of RACKs without affecting the proximity of PKCs to their substrates act as activators of PKC translocation and activity [127]. Second-generation peptide inhibitors may interfere with the interaction between PKC and a specific substrate on the same RACK [127]. The advantages of peptide drugs are their flexibility and naturally selected fit for a particular protein interaction site, which enable them to interact more effectively and specifically with proteins than rigid small molecules do and to interfere with multiple interaction sites on a protein [127]. Isoenzyme-specific peptide drugs such as PKC $\epsilon$ -activating and PKC $\delta$ -inhibiting peptides have been used as pharmacological modulators of PKC activity in basic research and animal models of disease, in particular improving outcomes in models of cardiovascular diseases and interventions [127,128].

### Pseudosubstrate peptides and ZIP ( $\zeta$ -inhibitory peptide)

ZIP (Table 1) is a myristoylated putative PKC $\zeta$ -inhibiting peptide composed of the pseudosubstrate sequence of PKC $\zeta$ . ZIP has been employed prominently in the past decade as the primary pharmacological tool with which Sacktor and co-workers have attempted to establish PKM $\zeta$ , an alternative transcript of PKC $\zeta$  that encodes its catalytic domain, as the kinase necessary and sufficient for the maintenance of LTP (long-term potentiation) of synaptic transmission and for learning and memory [95,96,129–131]. ZIP does indeed inhibit both PKC $\zeta$  and PKM $\zeta$  *in vitro* [23,95] and consistently abolishes LTP and various measures of memory in rats [95,96,129–131]. However, the effectiveness of ZIP in targeting PKM $\zeta$  in the vastly more complex environment of cells and tissues was never determined until 2012, when

cellular studies revealed that this peptide does not inhibit PKM $\zeta$  in mammalian cells or rat hippocampal brain slices in contexts where the positive control staurosporine does [23]. *In vitro*, the highly basic ZIP (and scrambled ZIP [23]) may inhibit PKM $\zeta$  by competing with the basic PKC substrate peptide for active-site occupation; however, in cells, the effective concentration of these basic peptides is considerably lower as a result of binding to any number of non-specific, negatively charged surfaces in a complex cellular environment. It is also possible that the pseudosubstrate sequence of PKC $\zeta$  binds with much higher affinity to a cellular target and that ZIP disrupts this interaction rather than the enzymatic activity of PKM $\zeta$ . Furthermore, in contrast with the regulatory mechanism of conventional PKCs, a recent study reports that the atypical C1 domain, rather than the pseudosubstrate of PKC $\zeta$ , is primarily responsible for allosterically autoinhibiting PKC $\zeta$  activity; deletion of the pseudosubstrate does not significantly increase the activity of the kinase [114]. Rather, the function of this highly positively charged pseudosubstrate is to interact with other regions on the protein to provide thermal stability; lipid-activated removal of the pseudosubstrate from the kinase active site results in loss of PKC $\zeta$  stability [114]. Recent reports that mice lacking PKM $\zeta$  have normal memory and learning, and continue to be sensitive to ZIP, confirm cellular pharmacological data that the target of ZIP is not PKM $\zeta$  [132,133]. Thus the cellular pharmacology of PKM $\zeta$  contrasts with its *in vitro* profile, underscoring the importance of establishing the cellular activity of pharmacological tools.

It is perhaps not surprising that pseudosubstrate-derived peptides are not effective PKC inhibitors in cells. First, the affinities of these peptides for PKC are several orders of magnitude lower than what is required for them to effectively bind PKC in cells. Peptides that effectively inhibit the related kinase PKA in cells bind with nanomolar affinity. PKI, the gold standard in inhibitory peptides that was originally characterized by John Scott and co-workers, binds the catalytic subunit of PKA with a  $K_d$  value of 4.8 nM [134]. Peptides such as Ht31 and AKAP-*IS*, which effectively disrupt localized PKA signalling by displacing PKA from AKAPs, bind with affinities of 2–4 nM [135]. In contrast, the affinities of pseudosubstrate peptides for PKC are in the micromolar range. Thus the field awaits the generation of higher-affinity peptides before peptides as a class can be applied as effective pharmacological inhibitors of PKC in cells. Secondly, although an intramolecular pseudosubstrate peptide may be able to autoinhibit a PKC isoenzyme, the isolated peptides may not be isoenzyme- or even PKC-specific inhibitors. Indeed, the pseudosubstrate peptide of PKC $\alpha$  is not a PKC-specific inhibitor, inhibiting also CaMKII and MLCK (myosin light-chain kinase) at micromolar concentrations *in vitro* [136]. Furthermore, on the basis of the unbiased screen of Cantley and co-workers for the optimal substrate sequences of individual PKC isoenzymes using an orientated peptide library, there is limited isoenzyme selectivity in substrate sequence recognition [137]. Thus, due to the demonstrated lack of specificity of pseudosubstrate-derived peptide inhibitors, results obtained using them should be interpreted with caution.

## DISCREDITED PKC INHIBITORS

### Rottlerin

Rottlerin (Table 1), also known as mallotoxin, was originally reported to be a PKC and CaMKIII inhibitor that inhibited PKC $\delta$  ( $IC_{50}$  = 3–6  $\mu$ M) with a 5–10-fold greater potency than conventional PKC isoenzymes and a 13–33-fold greater potency than other novel and atypical PKC isoenzymes tested *in vitro*

[138]. On the basis of this report, hundreds of published studies have used rottlerin to draw conclusions about the role of PKC $\delta$  in a variety of cellular processes. It has been over a decade, however, since rottlerin was debunked as a PKC $\delta$  inhibitor, with multiple independent investigators finding no effect of even 10–20  $\mu$ M rottlerin on PKC $\delta$  (or PKC $\alpha$  or PKC $\zeta$ ) activity *in vitro* [73,82,139]. Instead, rottlerin strongly inhibits many other protein kinases such as PRAK (p38-regulated/activated protein kinase) and MAPKAP-K2 [73] or such as CHK2, PLK1 (Polo-like kinase 1), PIM3 and SRPK1 [SRSF (serine/arginine-rich splicing factor) protein kinase 1] [82] much more potently *in vitro*. Possible explanations for this disparity include the use of the generic PKC substrate protamine in the original report, in contrast with the use of the optimal PKC $\delta$  substrate peptide in subsequent studies [139], and the presence of contaminants in the rottlerin used in the original report. Furthermore, treatment of cells with 10  $\mu$ M rottlerin does not inhibit phorbol ester-stimulated PKC $\delta$  activity, as measured using the isoenzyme-specific PKC $\delta$  activity reporter  $\delta$ CKAR, under conditions where 750 nM Gö6983 completely abolishes it [34]. Additionally, the cellular effects of rottlerin are not reproduced by the use of *bona fide* PKC inhibitors, by kinase-dead PKC $\delta$  or siRNA (small interfering RNA) against PKC $\delta$ , or by down-regulation of PKC $\delta$  with prolonged phorbol ester treatment [140]. Even more damningly, rottlerin continues to produce its effects in the absence of PKC $\delta$  activity (when inhibited by the *bona fide* PKC inhibitor BisI) [139] or in the absence of PKC $\delta$  protein (in cells from PKC $\delta$ -knockout mice or when down-regulated by phorbol esters) [140–143].

The widespread cellular effects of rottlerin on apoptosis and mitochondrial ROS (reactive oxygen species) production can be attributed to its activity as a mitochondrial uncoupler [139,140,144], a compound that uncouples ATP synthesis from respiration. Similar to the mitochondrial uncoupler FCCP (carbonyl cyanide *p*-trifluoromethoxyphenylhydrazone), which collapses the proton gradient across the mitochondrial inner membrane by acting as a protonophore, rottlerin depolarizes the mitochondrial membrane potential that drives ATP synthesis, thereby increasing the cellular oxygen consumption rate while reducing cellular ATP levels, activating AMP-dependent protein kinase and changing the phosphorylation of many signalling proteins [139,140]. Among these phosphorylation changes, tyrosine phosphorylation of PKC $\delta$ , which can activate PKC $\delta$  in certain contexts, can be indirectly blocked by rottlerin [139,140]. Although these effects are not mimicked by PKC inhibitors, they are mimicked by other mitochondrial uncouplers [139]. Responsible drug vendors should follow the lead of LC Labs, which has discontinued its sales of rottlerin, by at least ceasing to market rottlerin as a PKC $\delta$  inhibitor.

### Chelerythrine

Chelerythrine (Table 1) is a benzophenanthridine alkaloid that was originally reported in 1990 to be a potent *in vitro* substrate-competitive active-site inhibitor of PKC purified from rat brain, with an  $IC_{50}$  value of 0.66  $\mu$ M (at 100  $\mu$ M ATP), selectivity for PKC over PKA, CaMK and an unspecified tyrosine kinase, and anti-tumoural activity [145]. Since then, multiple independent investigators have debunked chelerythrine as a PKC inhibitor, both *in vitro* [73,76,146] and in cells [103]. In *in vitro* studies that contrast directly with the original report, chelerythrine did not inhibit either the basal or phorbol ester-induced activity of PKC purified from rat, mouse or calf brain, whereas staurosporine as a positive control did [146]. In fact, concentrations of chelerythrine up to 100  $\mu$ M enhanced the activity of PKC purified from rat and mouse brain [146]. At 500 nM, chelerythrine did not

inhibit any PKC isoenzyme (purified from insect cells) or any other kinase in a panel of 300 kinases assayed *in vitro* [76]. At 10  $\mu\text{M}$ , chelerythrine did not inhibit PKC $\alpha$  or any other protein kinase in a panel of 24 kinases assayed *in vitro* [73]. At 10  $\mu\text{M}$  or less, chelerythrine did not inhibit PKC $\zeta$  *in vitro* [147]. Because chelerythrine inhibited only lipid-stimulated and not basal PKC activity in the original report, we speculate that the positive charge of this amphipathic molecule may have been responsible for its reported effect by non-specifically reducing the surface charge of the anionic membranes needed as a cofactor in PKC activation. In cellular studies, concentrations of chelerythrine up to 10  $\mu\text{M}$  produced no effect on endogenous PKC in contexts where positive control PKC inhibitors did suppress PKC activity [103]. Furthermore, the cellular effects produced by chelerythrine have been found to be independent of PKC [148].

The continued marketing of chelerythrine as a specific PKC inhibitor has resulted in the inappropriate attribution of numerous and wide-ranging biological effects to PKC activity. Most prominently in the past decade, chelerythrine has been employed in the neurobiology field to implicate PKM $\zeta$  in long-term memory maintenance [95,130], even though the effectiveness of chelerythrine in inhibiting PKM $\zeta$  in the complex milieu of mammalian cells and tissues had never been established. In fact, it was recently demonstrated that chelerythrine does not inhibit PKM $\zeta$  in either mammalian cell lines or rat hippocampal brain slices [23]. Rather, chelerythrine is a cytotoxic and apoptosis-inducing drug [23,145,149,150] with mechanisms of action independent of PKC. For example, chelerythrine has been shown by NMR experiments to bind and inhibit the pro-survival Bcl-2 family member Bcl<sub>XL</sub> [150,151]. Furthermore, the imidium bond in chelerythrine can interact with the thiol groups of alanine aminotransferase [152]. The closely related benzophenanthradine alkaloid sanguinarine has also been shown to bind Bcl<sub>XL</sub> [151], and its imidium bond has also been shown to react with the nucleophilic and anionic moieties of amino acids and to intercalate into DNA [151,153].

## GENETIC MANIPULATION OF PKC

### Kinase-dead mutants

As a complementary or alternative approach to pharmacological inhibition, studies frequently make use of kinase-dead PKC constructs to address the role of PKC activity in biological processes. These kinase-dead constructs are all generated by mutating one of a number of conserved residues in the active site of PKC, which renders the mutant kinase itself essentially inactive and potentially also generates a dominant-negative effect in cells from competition against endogenous PKC [154]. However, such mutations often compromise phosphorylation of the priming sites on PKC. Thus priming phosphorylations of mutants should be verified, and effects obtained using unprimed mutants should be treated with caution, as they may be the artifactual result of improper protein conformation, rather than of loss of kinase activity. Furthermore, most kinase-dead mutations reduce activity significantly (10–100-fold), but do not entirely abolish it. As a consequence, some 'kinase-dead' constructs can still autophosphorylate their C-terminal priming sites. Kinase-dead PKCs that remain unphosphorylated are typically degraded or may localize to a different spatial compartment than processed PKCs, potentially producing off-target effects.

The most commonly mutated residue reported in the literature to generate a kinase-dead PKC corresponds to Lys<sup>376</sup> of mouse PKC $\delta$  and is mutated either to methionine or arginine. This

positively charged lysine residue hydrogen bonds with the  $\alpha$  and  $\beta$  phosphates of ATP and forms an ion pair with a conserved glutamate in the C-helix that stabilizes the N-lobe of the kinase domain [102]. Thus mutation of this residue inhibits activity by impairing phosphoryl transfer.

Another conserved residue that has been mutated to generate a kinase-dead PKC corresponds to Asp<sup>471</sup> of mouse PKC $\delta$  and is mutated to either alanine or asparagine. This aspartate residue acts as the catalytic base to abstract the proton of the hydroxy group of the serine or threonine residue on the substrate to co-ordinate it for phosphorylation [102]. Mutation of this residue has been reported to produce no loss of priming phosphorylations on PKC $\epsilon$  and PKC $\alpha$  [102]. However, although this mutant in PKC $\epsilon$  loses all *in vitro* kinase activity [102], the naturally occurring substitution of asparagine for the catalytic aspartate in the pseudokinase ErbB3 preserves enough residual kinase activity for ErbB3 to autophosphorylate [155].

A third conserved residue that was recently mutated to inactivate a kinase corresponds to Ala<sup>374</sup> of mouse PKC $\delta$  and was originally mutated in KSR1 (kinase suppressor of Ras 1) to a phenylalanine [156]. This alanine residue in the N-lobe of kinases forms part of a three-dimensional catalytic hydrophobic spine that stabilizes the active conformation of a kinase and interacts with the top of the adenine ring in ATP [156]. According to molecular modelling of CRAF, this alanine to phenylalanine mutation may stabilize the catalytic hydrophobic spine to maintain an active conformation while blocking ATP binding, creating a pseudokinase [156]. Indeed, the KSR1 alanine/phenylalanine mutant does not bind ATP and is not active (it cannot enable RasV12 transformation-induced colony formation), but retains its scaffolding of BRAF and CRAF [156].

### Gatekeeper mutants and chemical genetics

Chemical genetics approaches to kinase biology, pioneered by the laboratory of Kevan Shokat in the late 1990s, afford an elegant approach to interrogate the role played by the intrinsic catalytic activity of a particular PKC isoenzyme [157–160]. Specifically, the genetic engineering of protein kinases to preferentially bind to co-engineered small-molecule ligands provides a method to specifically target the kinase of interest. These ligands are either modified nucleotides that can be used to radiolabel and thus identify the substrates specifically phosphorylated by the engineered kinase or small-molecule inhibitors derived from a PP (pyrazolo[3,4-D]pyrimidine) inhibitor of Src family kinases that specifically inhibit the engineered kinase. The chemical genetics approach to identifying kinase-specific substrates was made possible by the discovery that, when a certain bulky conserved hydrophobic residue in the kinase active site that interacts with the adenine N<sup>6</sup> amino group on ATP is mutated to a smaller alanine or glycine, this mutation slightly enlarges the active site to accommodate a modified bulkier ATP that cannot bind to the active site of wild-type kinases. This critical residue has become known as the gatekeeper residue for its role in guarding the active site of wild-type kinases against the entry of engineered ligands and allowing their entry in gatekeeper mutants. The chemical genetics approach to inhibiting a specific kinase quickly followed when mutation of the same bulky residue was found to accommodate the binding of cell-permeable mutant-selective PP-derived small-molecule inhibitors, whose N<sup>4</sup> position corresponds to the N<sup>6</sup> position of ATP. This chemical genetics approach was recently employed for mechanistic studies to demonstrate a role for the intrinsic catalytic activity of PKC $\delta$  in its isoenzyme-specific interaction with mitochondria [34], as well as for an

organism-based screen to identify cellular substrates of PKC $\epsilon$  [161].

### Genetic deletion or depletion of PKC

PKC isoenzymes, in general, have relatively long half-lives, so effective knockdown by siRNA can be challenging [68,162,163]. However, with the exception of PKC $\epsilon$ , knockout of PKC isoenzymes results in viable mice (e.g. [133,164–168]). Thus analysis of mouse embryonic fibroblasts from isoenzyme-specific knockout mice provides a helpful tool for examining isoenzyme-specific function (e.g. [16,169]).

### CONCLUSION AND PERSPECTIVES

We have aimed in the present review to clarify the current status and mechanisms of action of the *bona fide* and discredited pharmacological tools in the PKC toolbox. A wide range of compounds in the forms of activators, small-molecule inhibitors and peptides have emerged to modulate the catalytic activity, subcellular distribution, protein interactions and life cycle of PKC. Coupled with the advent of FRET-based reporters to examine the spatiotemporal dynamics of PKC signalling, tools should no longer limit our accurate assessment of the biological function of PKC. Given the importance of time and space in cell signalling, future advances in developing tools that reversibly turn on or off specific isoenzymes at specific cellular locations, as well as tools that allow supra-high definition of PKC subcellular locations, will be welcome additions to the PKC toolbox. In this regard, novel and light-sensitive tags such as the recently described mini-SOG (a 106 amino acid tag whose light-dependent generation of singlet oxygen affords a way to locally inactivate proteins and to visualize proteins with high-resolution electron microscopy) holds promise for defining the precise cellular location of specific isoenzymes [170]. In the meantime, it is our hope that, in refining the PKC pharmacological toolbox, we will assist investigators in selecting the best tools for studying the role of PKC in their systems and minimize the formation of erroneous conclusions from the use of non-specific compounds.

### ACKNOWLEDGEMENTS

We thank members of the Newton laboratory for helpful discussions and a critical reading of this paper.

### FUNDING

Work in the authors' laboratory was supported by the National Institutes of Health [grant numbers GM43154 and P01 DK054441 (to A.C.N.)] and in part by the University of California, San Diego Graduate Training Program in Cellular and Molecular Pharmacology through the National Institute of General Medical Sciences [grant number T32 GM007752 (to A.X.W.)].

### REFERENCES

- Castagna, M., Takai, Y., Kaibuchi, K., Sano, K., Kikkawa, U. and Nishizuka, Y. (1982) Direct activation of calcium-activated, phospholipid-dependent protein kinase by tumor-promoting phorbol esters. *J. Biol. Chem.* **257**, 7847–7851
- Newton, A. C. (2004) Diacylglycerol's affair with protein kinase C turns 25. *Trends Pharmacol. Sci.* **25**, 175–177
- Newton, A. C. (2010) Protein kinase C: poised to signal. *Am. J. Physiol. Endocrinol. Metab.* **298**, E395–E402
- Griner, E. M. and Kazanietz, M. G. (2007) Protein kinase C and other diacylglycerol effectors in cancer. *Nat. Rev. Cancer* **7**, 281–294
- Reyland, M. E. (2009) Protein kinase C isoforms: multi-functional regulators of cell life and death. *Front. Biosci.* **14**, 2386–2399
- Rosse, C., Linch, M., Kermorgant, S., Cameron, A. J., Boeckeler, K. and Parker, P. J. (2010) PKC and the control of localized signal dynamics. *Nat. Rev. Mol. Cell Biol.* **11**, 103–112
- Steinberg, S. F. (2008) Structural basis of protein kinase C isoform function. *Physiol. Rev.* **88**, 1341–1378
- Newton, A. C. (2001) Protein kinase C: structural and spatial regulation by phosphorylation, cofactors, and macromolecular interactions. *Chem. Rev.* **101**, 2353–2364
- Newton, A. C. (2009) Lipid activation of protein kinases. *J. Lipid Res.* **50** (Suppl.), S266–S271
- Giorgione, J. R., Lin, J. H., McCammon, J. A. and Newton, A. C. (2006) Increased membrane affinity of the C1 domain of protein kinase C $\delta$  compensates for the lack of involvement of its C2 domain in membrane recruitment. *J. Biol. Chem.* **281**, 1660–1669
- Dries, D. R., Gallegos, L. L. and Newton, A. C. (2007) A single residue in the C1 domain sensitizes novel protein kinase C isoforms to cellular diacylglycerol production. *J. Biol. Chem.* **282**, 826–830
- Hernandez, A. I., Blace, N., Cray, J. F., Serrano, P. A., Leitges, M., Libien, J. M., Weinstein, G., Tcherapanov, A. and Sacktor, T. C. (2003) Protein kinase M $\zeta$  synthesis from a brain mRNA encoding an independent protein kinase C $\zeta$  catalytic domain. Implications for the molecular mechanism of memory. *J. Biol. Chem.* **278**, 40305–40316
- Jiang, K., Apostolatos, A. H., Ghansah, T., Watson, J. E., Vickers, T., Cooper, D. R., Epling-Burnette, P. K. and Patel, N. A. (2008) Identification of a novel antiapoptotic human protein kinase C $\delta$  isoform, PKC $\delta$ VIII in NT2 cells. *Biochemistry* **47**, 787–797
- Ueyama, T., Ren, Y., Ohmori, S., Sakai, K., Tamaki, N. and Saito, N. (2000) cDNA cloning of an alternative splicing variant of protein kinase C $\delta$  (PKC $\delta$ III), a new truncated form of PKC $\delta$ , in rats. *Biochem. Biophys. Res. Commun.* **269**, 557–563
- Kim, J. D., Seo, K. W., Lee, E. A., Quang, N. N., Cho, H. R. and Kwon, B. (2011) A novel mouse PKC $\delta$  splice variant, PKC $\delta$ IX, inhibits etoposide-induced apoptosis. *Biochem. Biophys. Res. Commun.* **410**, 177–182
- O'Neill, A. K., Gallegos, L. L., Justilien, V., Garcia, E. L., Leitges, M., Fields, A. P., Hall, R. A. and Newton, A. C. (2011) Protein kinase C $\alpha$  promotes cell migration through a PDZ-dependent interaction with its novel substrate discs large homolog 1 (DLG1). *J. Biol. Chem.* **286**, 43559–43568
- Sheng, M. and Sala, C. (2001) PDZ domains and the organization of supramolecular complexes. *Annu. Rev. Neurosci.* **24**, 1–29
- Newton, A. C. (2003) Regulation of the ABC kinases by phosphorylation: protein kinase C as a paradigm. *Biochem. J.* **370**, 361–371
- Gould, C. M. and Newton, A. C. (2008) The life and death of protein kinase C. *Curr. Drug Targets* **9**, 614–625
- Gallegos, L. L. and Newton, A. C. (2008) Spatiotemporal dynamics of lipid signaling: protein kinase C as a paradigm. *IUBMB Life* **60**, 782–789
- Roffey, J., Rosse, C., Linch, M., Hibbert, A., McDonald, N. Q. and Parker, P. J. (2009) Protein kinase C intervention: the state of play. *Curr. Opin. Cell Biol.* **21**, 268–279
- Das Evcimen, N. and King, G. L. (2007) The role of protein kinase C activation and the vascular complications of diabetes. *Pharmacol. Res.* **55**, 498–510
- Wu-Zhang, A. X., Schramm, C. L., Nabavi, S., Malinow, R. and Newton, A. C. (2012) Cellular pharmacology of protein kinase M $\zeta$  (PKM $\zeta$ ) contrasts with its *in vitro* profile: implications for PKM $\zeta$  as a mediator of memory. *J. Biol. Chem.* **287**, 12879–12885
- Cohen, P. (2010) Guidelines for the effective use of chemical inhibitors of protein function to understand their roles in cell regulation. *Biochem. J.* **425**, 53–54
- Violin, J. D. and Newton, A. C. (2003) Pathway illuminated: visualizing protein kinase C signaling. *IUBMB Life* **55**, 653–660
- Violin, J. D., Zhang, J., Tsien, R. Y. and Newton, A. C. (2003) A genetically encoded fluorescent reporter reveals oscillatory phosphorylation by protein kinase C. *J. Cell Biol.* **161**, 899–909
- Ting, A. Y., Kain, K. H., Klemke, R. L. and Tsien, R. Y. (2001) Genetically encoded fluorescent reporters of protein tyrosine kinase activities in living cells. *Proc. Natl. Acad. Sci. U.S.A.* **98**, 15003–15008
- Zhang, J., Ma, Y., Taylor, S. S. and Tsien, R. Y. (2001) Genetically encoded reporters of protein kinase A activity reveal impact of substrate tethering. *Proc. Natl. Acad. Sci. U.S.A.* **98**, 14997–15002
- Kunkel, M. T., Ni, Q., Tsien, R. Y., Zhang, J. and Newton, A. C. (2005) Spatio-temporal dynamics of protein kinase B/Akt signaling revealed by a genetically encoded fluorescent reporter. *J. Biol. Chem.* **280**, 5581–5587
- Kunkel, M. T., Toker, A., Tsien, R. Y. and Newton, A. C. (2007) Calcium-dependent regulation of protein kinase D revealed by a genetically encoded kinase activity reporter. *J. Biol. Chem.* **282**, 6733–6742
- Kunkel, M. T. and Newton, A. C. (2009) Spatiotemporal dynamics of kinase signaling visualized by targeted reporters. *Curr. Protoc. Chem. Biol.* **1**, 17–18

- 32 Scott, J. D. and Newton, A. C. (2012) Shedding light on local kinase activation. *BMC Biol.* **10**, 61
- 33 Kajimoto, T., Sawamura, S., Tohyama, Y., Mori, Y. and Newton, A. C. (2010) Protein kinase C $\delta$ -specific activity reporter reveals agonist-evoked nuclear activity controlled by Src family of kinases. *J. Biol. Chem.* **285**, 41896–41910
- 34 Wu-Zhang, A. X., Murphy, A. N., Bachman, M. and Newton, A. C. (2012) Isozyme-specific interaction of protein kinase C $\delta$  (PKC $\delta$ ) with mitochondria dissected using live cell fluorescence imaging. *J. Biol. Chem.* **287**, 37891–37906
- 35 Gadi, D., Wagenknecht-Wiesner, A., Holowka, D. and Baird, B. (2011) Sequestration of phosphoinositides by mutated MARCKS effector domain inhibits stimulated Ca<sup>2+</sup> mobilization and degranulation in mast cells. *Mol. Biol. Cell* **22**, 4908–4917
- 36 Gallegos, L. L., Kunkel, M. T. and Newton, A. C. (2006) Targeting protein kinase C activity reporter to discrete intracellular regions reveals spatiotemporal differences in agonist-dependent signaling. *J. Biol. Chem.* **281**, 30947–30956
- 37 Kunkel, M. T., Garcia, E. L., Kajimoto, T., Hall, R. A. and Newton, A. C. (2009) The protein scaffold NHERF-1 controls the amplitude and duration of localized protein kinase D activity. *J. Biol. Chem.* **284**, 24653–24661
- 38 Hoshi, N., Langeberg, L. K., Gould, C. M., Newton, A. C. and Scott, J. D. (2010) Interaction with AKAP79 modifies the cellular pharmacology of PKC. *Mol. Cell* **37**, 541–550
- 39 Kunkel, M. T. and Newton, A. C. (2010) Calcium transduces plasma membrane receptor signals to produce diacylglycerol at Golgi membranes. *J. Biol. Chem.* **285**, 22748–22752
- 40 Palmer, A. E., Giacomello, M., Kortemme, T., Hires, S. A., Lev-Ram, V., Baker, D. and Tsien, R. Y. (2006) Ca<sup>2+</sup> indicators based on computationally redesigned calmodulin-peptide pairs. *Chem. Biol.* **13**, 521–530
- 41 Sato, M., Ueda, Y. and Umezawa, Y. (2006) Imaging diacylglycerol dynamics at organelle membranes. *Nat. Methods* **3**, 797–799
- 42 Tewson, P., Westenberg, M., Zhao, Y., Campbell, R. E., Quinn, A. M. and Hughes, T. E. (2012) Simultaneous detection of Ca<sup>2+</sup> and diacylglycerol signaling in living cells. *PLoS ONE* **7**, e42791
- 43 Blumberg, P. M., Kedei, N., Lewin, N. E., Yang, D., Tao, J., Telek, A. and Geczy, T. (2010) Phorbol esters and diacylglycerol: the PKC activators. In *Protein Kinase C in Cancer Signaling and Therapy* (Kazanietz, M. G., ed.), pp. 25–53, Springer, New York
- 44 Mosior, M. and Newton, A. C. (1996) Calcium-independent binding to interfacial phorbol esters causes protein kinase C to associate with membranes in the absence of acidic lipids. *Biochemistry* **35**, 1612–1623
- 45 Sharkey, N. A., Leach, K. L. and Blumberg, P. M. (1984) Competitive inhibition by diacylglycerol of specific phorbol ester binding. *Proc. Natl. Acad. Sci. U.S.A.* **81**, 607–610
- 46 Kikkawa, U., Takai, Y., Tanaka, Y., Miyake, R. and Nishizuka, Y. (1983) Protein kinase C as a possible receptor protein of tumor-promoting phorbol esters. *J. Biol. Chem.* **258**, 11442–11445
- 47 Kazanietz, M. G. (2002) Novel “non-kinase” phorbol ester receptors: the C1 domain connection. *Mol. Pharmacol.* **61**, 1–9
- 48 Hecker, E. (1968) Cocarcinogenic principles from the seed oil of *Croton tiglium* and from other Euphorbiaceae. *Cancer Res.* **28**, 2338–2349
- 49 Driedger, P. E. and Blumberg, P. M. (1980) Specific binding of phorbol ester tumor promoters. *Proc. Natl. Acad. Sci. U.S.A.* **77**, 567–571
- 50 Zhang, G., Kazanietz, M. G., Blumberg, P. M. and Hurley, J. H. (1995) Crystal structure of the cys2 activator-binding domain of protein kinase C $\delta$  in complex with phorbol ester. *Cell* **81**, 917–924
- 51 Dries, D. R. and Newton, A. C. (2008) Kinetic analysis of the interaction of the C1 domain of protein kinase C with lipid membranes by stopped-flow spectroscopy. *J. Biol. Chem.* **283**, 7885–7893
- 52 Zayed, S., Sorg, B. and Hecker, E. (1984) Structure activity relations of polyfunctional diterpenes of the tigliane type, VI1. *Planta Med.* **50**, 65–69
- 53 Furstemberger, G. and Hecker, E. (1972) Mode of action of carcinogenic plant components. *Planta Med.* **22**, 241–266
- 54 Wang, Q. J., Fang, T. W., Fenick, D., Garfield, S., Bienfait, B., Marquez, V. E. and Blumberg, P. M. (2000) The lipophilicity of phorbol esters as a critical factor in determining the pattern of translocation of protein kinase C $\delta$  fused to green fluorescent protein. *J. Biol. Chem.* **275**, 12136–12146
- 55 Marquez, V. E. and Blumberg, P. M. (2003) Synthetic diacylglycerols (DAG) and DAG-lactones as activators of protein kinase C (PK-C). *Acc. Chem. Res.* **36**, 434–443
- 56 Tran, P. L. and Deugnier, M. A. (1985) Intracellular localization of 12-O-3-N-dansylamino TPA in C3H/10T1/2 mouse cell line. *Carcinogenesis* **6**, 433–439
- 57 Slater, S. J., Ho, C. and Stubbs, C. D. (2002) The use of fluorescent phorbol esters in studies of protein kinase C-membrane interactions. *Chem. Phys. Lipids* **116**, 75–91
- 58 Braun, D. C., Cao, Y., Wang, S., Garfield, S. H., Hur, G. M. and Blumberg, P. M. (2005) Role of phorbol ester localization in determining protein kinase C or RasGRP3 translocation: real-time analysis using fluorescent ligands and proteins. *Mol. Cancer Ther.* **4**, 141–150
- 59 Fallen, R. S. and Gooderham, M. (2012) Ingenol mebutate: an introduction. *Skin Therapy Lett.* **17**, 1–3
- 60 Shao, L., Lewin, N. E., Lorenzo, P. S., Hu, Z., Enyedy, I. J., Garfield, S. H., Stone, J. C., Marner, F. J., Blumberg, P. M. and Wang, S. (2001) Iridals are a novel class of ligands for phorbol ester receptors with modest selectivity for the RasGRP receptor subfamily. *J. Med. Chem.* **44**, 3872–3880
- 61 Pettit, G. R., Herald, C. L., Doubek, D. L., Herald, D. L., Arnold, E. and Clardy, J. (1982) Isolation and structure of bryostatin 1. *J. Am. Chem. Soc.* **104**, 6846–6848
- 62 Keck, G. E., Poudel, Y. B., Welch, D. S., Kraft, M. B., Truong, A. P., Stephens, J. C., Kedei, N., Lewin, N. E. and Blumberg, P. M. (2009) Substitution on the A-ring confers to bryopyran analogues the unique biological activity characteristic of bryostatins and distinct from that of the phorbol esters. *Org. Lett.* **11**, 593–596
- 63 Kortmansky, J. and Schwartz, G. K. (2003) Bryostatin-1: a novel PKC inhibitor in clinical development. *Cancer Invest.* **21**, 924–936
- 64 Mayer, A. M., Glaser, K. B., Cuevas, C., Jacobs, R. S., Kem, W., Little, R. D., McIntosh, J. M., Newman, D. J., Potts, B. C. and Shuster, D. E. (2010) The odyssey of marine pharmaceuticals: a current pipeline perspective. *Trends Pharmacol. Sci.* **31**, 255–265
- 65 Szallasi, Z., Denning, M. F., Smith, C. B., Dlugosz, A. A., Yuspa, S. H., Pettit, G. R. and Blumberg, P. M. (1994) Bryostatin 1 protects protein kinase C- $\delta$  from down-regulation in mouse keratinocytes in parallel with its inhibition of phorbol ester-induced differentiation. *Mol. Pharmacol.* **46**, 840–850
- 66 Szallasi, Z., Smith, C. B., Pettit, G. R. and Blumberg, P. M. (1994) Differential regulation of protein kinase C isozymes by bryostatin 1 and phorbol 12-myristate 13-acetate in NIH 3T3 fibroblasts. *J. Biol. Chem.* **269**, 2118–2124
- 67 Wang, Q. J., Bhattacharyya, D., Garfield, S., Nacro, K., Marquez, V. E. and Blumberg, P. M. (1999) Differential localization of protein kinase C  $\delta$  by phorbol esters and related compounds using a fusion protein with green fluorescent protein. *J. Biol. Chem.* **274**, 37233–37239
- 68 von Burstin, V. A., Xiao, L. and Kazanietz, M. G. (2010) Bryostatin 1 inhibits phorbol ester-induced apoptosis in prostate cancer cells by differentially modulating protein kinase C (PKC)  $\delta$  translocation and preventing PKC $\delta$ -mediated release of tumor necrosis factor- $\alpha$ . *Mol. Pharmacol.* **78**, 325–332
- 69 Kedei, N., Lubart, E., Lewin, N. E., Telek, A., Lim, L., Mannan, P., Garfield, S. H., Kraft, M. B., Keck, G. E., Kolusheva, S. et al. (2011) Some phorbol esters might partially resemble bryostatin 1 in their actions on LNCaP prostate cancer cells and U937 leukemia cells. *ChemBioChem* **12**, 1242–1251
- 70 Wender, P. A., Hinkle, K. W., Koehler, M. F. and Lipka, B. (1999) The rational design of potential chemotherapeutic agents: synthesis of bryostatin analogues. *Med. Res. Rev.* **19**, 388–407
- 71 Keck, G. E., Kraft, M. B., Truong, A. P., Li, W., Sanchez, C. C., Kedei, N., Lewin, N. E. and Blumberg, P. M. (2008) Convergent assembly of highly potent analogues of bryostatin 1 via pyran annulation: bryostatin look-alikes that mimic phorbol ester function. *J. Am. Chem. Soc.* **130**, 6660–6661
- 72 Wender, P. A., Verma, V. A., Paxton, T. J. and Pillow, T. H. (2008) Function-oriented synthesis, step economy, and drug design. *Acc. Chem. Res.* **41**, 40–49
- 73 Davies, S. P., Reddy, H., Caivano, M. and Cohen, P. (2000) Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem. J.* **351**, 95–105
- 74 Karaman, M. W., Herrgard, S., Treiber, D. K., Gallant, P., Atteridge, C. E., Campbell, B. T., Chan, K. W., Ciceri, P., Davis, M. I., Edeen, P. T. et al. (2008) A quantitative analysis of kinase inhibitor selectivity. *Nat. Biotechnol.* **26**, 127–132
- 75 Davis, M. I., Hunt, J. P., Herrgard, S., Ciceri, P., Wodicka, L. M., Pallares, G., Hocker, M., Treiber, D. K. and Zarrinkar, P. P. (2011) Comprehensive analysis of kinase inhibitor selectivity. *Nat. Biotechnol.* **29**, 1046–1051
- 76 Anastassiadis, T., Deacon, S. W., Devarajan, K., Ma, H. and Peterson, J. R. (2011) Comprehensive assay of kinase catalytic activity reveals features of kinase inhibitor selectivity. *Nat. Biotechnol.* **29**, 1039–1045
- 77 Inoue, M., Kishimoto, A., Takai, Y. and Nishizuka, Y. (1977) Studies on a cyclic nucleotide-independent protein kinase and its proenzyme in mammalian tissues. *J. Biol. Chem.* **252**, 7610–7616
- 78 Martiny-Baron, G., Kazanietz, M. G., Mischak, H., Blumberg, P. M., Kochs, G., Hug, H., Marme, D. and Schachtele, C. (1993) Selective inhibition of protein kinase C isozymes by the indolocarbazole Gö 6976. *J. Biol. Chem.* **268**, 9194–9197
- 79 Gschwendt, M., Dieterich, S., Rennecke, J., Kittstein, W., Mueller, H. J. and Johannes, F. J. (1996) Inhibition of protein kinase C $\mu$  by various inhibitors. Differentiation from protein kinase C isoenzymes. *FEBS Lett.* **392**, 77–80
- 80 Grodsky, N., Li, Y., Bouzida, D., Love, R., Jensen, J., Nodes, B., Nonomiya, J. and Grant, S. (2006) Structure of the catalytic domain of human protein kinase C $\beta$  II complexed with a bisindolylmaleimide inhibitor. *Biochemistry* **45**, 13970–13981
- 81 Wilkinson, S. E., Parker, P. J. and Nixon, J. S. (1993) Isoenzyme specificity of bisindolylmaleimides, selective inhibitors of protein kinase C. *Biochem. J.* **294**, 335–337

- 82 Bain, J., Plater, L., Elliott, M., Spiro, N., Hastie, C. J., McLauchlan, H., Klevernic, I., Arthur, J. S., Alessi, D. R. and Cohen, P. (2007) The selectivity of protein kinase inhibitors: a further update. *Biochem. J.* **408**, 297–315
- 83 Alessi, D. R. (1997) The protein kinase C inhibitors Ro 318220 and GF 109203X are equally potent inhibitors of MAPKAP kinase-1 $\beta$  (Rsk-2) and p70 S6 kinase. *FEBS Lett.* **402**, 121–123
- 84 Evenou, J. P., Wagner, J., Zenke, G., Brinkmann, V., Wagner, K., Kovarik, J., Welzenbach, K. A., Weitz-Schmidt, G., Guntermann, C., Towbin, H. et al. (2009) The potent protein kinase C-selective inhibitor AEB071 (sotrastaurin) represents a new class of immunosuppressive agents affecting early T-cell activation. *J. Pharmacol. Exp. Ther.* **330**, 792–801
- 85 Wagner, J., von Matt, P., Sedrani, R., Albert, R., Cooke, N., Ehrhardt, C., Geiser, M., Rummel, G., Stark, W., Strauss, A. et al. (2009) Discovery of 3-(1H-indol-3-yl)-4-[2-(4-methylpiperazin-1-yl)quinazolin-4-yl]pyrrolo-2,5-dione (AEB071), a potent and selective inhibitor of protein kinase C isotypes. *J. Med. Chem.* **52**, 6193–6196
- 86 Skvara, H., Dawid, M., Kleyn, E., Wolff, B., Meingassner, J. G., Knight, H., Dumortier, T., Kopp, T., Fallahi, N., Stary, G. et al. (2008) The PKC inhibitor AEB071 may be a therapeutic option for psoriasis. *J. Clin. Invest.* **118**, 3151–3159
- 87 Jirousek, M. R., Gillig, J. R., Gonzalez, C. M., Heath, W. F., McDonald, 3rd, J. H., Neel, D. A., Rito, C. J., Singh, U., Stramm, L. E., Melikian-Badalian, A. et al. (1996) (S)-13-[(dimethylamino)methyl]-10,11,14,15-tetrahydro-4,9:16, 21-dimetheno-1H, 13H-dibenzo[e,k]pyrrolo[3,4-h][1,4,13]oxadiazacyclohexadecene-1,3(2H)-dione (LY333531) and related analogues: isozyme selective inhibitors of protein kinase C $\beta$ . *J. Med. Chem.* **39**, 2664–2671
- 88 Tanaka, M., Sagawa, S., Hoshi, J., Shimoma, F., Matsuda, I., Sakoda, K., Sasase, T., Shindo, M. and Inaba, T. (2004) Synthesis of anilino-monoindolylmaleimides as potent and selective PKC $\beta$  inhibitors. *Bioorg. Med. Chem. Lett.* **14**, 5171–5174
- 89 Graff, J. R., McNulty, A. M., Hanna, K. R., Konicek, B. W., Lynch, R. L., Bailey, S. N., Banks, C., Capen, A., Goode, R., Lewis, J. E. et al. (2005) The protein kinase C $\beta$ -selective inhibitor, Enzastaurin (LY317615.HCl), suppresses signaling through the AKT pathway, induces apoptosis, and suppresses growth of human colon cancer and glioblastoma xenografts. *Cancer Res.* **65**, 7462–7469
- 90 Green, L. J., Marder, P., Ray, C., Cook, C. A., Jaken, S., Musib, L. C., Herbst, R. S., Carducci, M., Britten, C. D., Basche, M. et al. (2006) Development and validation of a drug activity biomarker that shows target inhibition in cancer patients receiving enzastaurin, a novel protein kinase C- $\beta$  inhibitor. *Clin. Cancer Res.* **12**, 3408–3415
- 91 Ikenoue, T., Inoki, K., Yang, Q., Zhou, X. and Guan, K. L. (2008) Essential function of TORC2 in PKC and Akt turn motif phosphorylation, maturation and signalling. *EMBO J.* **27**, 1919–1931
- 92 Omura, S., Iwai, Y., Hirano, A., Nakagawa, A., Awaya, J., Tsuchiya, H., Takahashi, Y. and Masuma, R. (1977) A new alkaloid AM-2282 of streptomycetes origin. Taxonomy, fermentation, isolation and preliminary characterization. *J. Antibiot.* **30**, 275–282
- 93 Tamaoki, T., Nomoto, H., Takahashi, I., Kato, Y., Morimoto, M. and Tomita, F. (1986) Staurosporine, a potent inhibitor of phospholipid/Ca<sup>++</sup> dependent protein kinase. *Biochem. Biophys. Res. Commun.* **135**, 397–402
- 94 Seynaeve, C. M., Kazanietz, M. G., Blumberg, P. M., Sausville, E. A. and Worland, P. J. (1994) Differential inhibition of protein kinase C isozymes by UCN-01, a staurosporine analogue. *Mol. Pharmacol.* **45**, 1207–1214
- 95 Ling, D. S., Benardo, L. S., Serrano, P. A., Blace, N., Kelly, M. T., Cray, J. F. and Sacktor, T. C. (2002) Protein kinase M $\zeta$  is necessary and sufficient for LTP maintenance. *Nat. Neurosci.* **5**, 295–296
- 96 Pastalkova, E., Serrano, P., Pinkhasova, D., Wallace, E., Fenton, A. A. and Sacktor, T. C. (2006) Storage of spatial information by the maintenance mechanism of LTP. *Science* **313**, 1141–1144
- 97 Dutil, E. M., Tokar, A. and Newton, A. C. (1998) Regulation of conventional protein kinase C isozymes by phosphoinositide-dependent kinase 1 (PKD-1). *Curr. Biol.* **8**, 1366–1375
- 98 Le Good, J. A., Ziegler, W. H., Parekh, D. B., Alessi, D. R., Cohen, P. and Parker, P. J. (1998) Protein kinase C isotypes controlled by phosphoinositide 3-kinase through the protein kinase PDK1. *Science* **281**, 2042–2045
- 99 Chou, M. M., Hou, W., Johnson, J., Graham, L. K., Lee, M. H., Chen, C. S., Newton, A. C., Schaffhausen, B. S. and Tokar, A. (1998) Regulation of protein kinase C $\zeta$  by PI 3-kinase and PDK-1. *Curr. Biol.* **8**, 1069–1077
- 100 Komander, D., Kular, G. S., Bain, J., Elliott, M., Alessi, D. R. and van Aalten, D. M. (2003) Structural basis for UCN-01 (7-hydroxystaurosporine) specificity and PDK1 (3-phosphoinositide-dependent protein kinase-1) inhibition. *Biochem. J.* **375**, 255–262
- 101 Kjær, S., Linch, M., Purkiss, A., Kostecky, B., Knowles, P. P., Rosse, C., Riou, P., Soudy, C., Kaye, S., Patel, B. et al. (2013) Adenosine-binding motif mimicry and cellular effects of a thieno[2,3-D]pyrimidine-based chemical inhibitor of atypical protein kinase C isozymes. *Biochem. J.* **451**, 329–342
- 102 Cameron, A. J., Escribano, C., Saurin, A. T., Kostecky, B. and Parker, P. J. (2009) PKC maturation is promoted by nucleotide pocket occupation independently of intrinsic kinase activity. *Nat. Struct. Mol. Biol.* **16**, 624–630
- 103 Gould, C. M., Antal, C. E., Reyes, G., Kunkel, M. T., Adams, R. A., Ziyar, A., Riveros, T. and Newton, A. C. (2011) Active site inhibitors protect protein kinase C from dephosphorylation and stabilize its mature form. *J. Biol. Chem.* **286**, 28922–28930
- 104 Okuzumi, T., Fiedler, D., Zhang, C., Gray, D. C., Aizenstein, B., Hoffman, R. and Shokat, K. M. (2009) Inhibitor hijacking of Akt activation. *Nat. Chem. Biol.* **5**, 484–493
- 105 Chan, T. O., Zhang, J., Rodeck, U., Pascal, J. M., Armen, R. S., Spring, M., Dumitru, C. D., Myers, V., Li, X., Cheung, J. Y. and Feldman, A. M. (2011) Resistance of Akt kinases to dephosphorylation through ATP-dependent conformational plasticity. *Proc. Natl. Acad. Sci. U.S.A.* **108**, E1120–E1127
- 106 Lin, K., Lin, J., Wu, W. I., Ballard, J., Lee, B. B., Gloor, S. L., Vigers, G. P., Morales, T. H., Friedman, L. S., Skelton, N. and Brandhuber, B. J. (2012) An ATP-site on-off switch that restricts phosphatase accessibility of Akt. *Sci. Signaling* **5**, ra37
- 107 Dutil, E. M., Keranen, L. M., DePaoli-Roach, A. A. and Newton, A. C. (1994) *In vivo* regulation of protein kinase C by trans-phosphorylation followed by autophosphorylation. *J. Biol. Chem.* **269**, 29359–29362
- 108 Dutil, E. M. and Newton, A. C. (2000) Dual role of pseudosubstrate in the coordinated regulation of protein kinase C by phosphorylation and diacylglycerol. *J. Biol. Chem.* **275**, 10697–10701
- 109 Akamine, P., Madhusudan, Wu, J., Xuong, N. H., Ten Eyck, L. F. and Taylor, S. S. (2003) Dynamic features of cAMP-dependent protein kinase revealed by apoenzyme crystal structure. *J. Mol. Biol.* **327**, 159–171
- 110 Narayana, N., Diller, T. C., Koide, K., Bunnage, M. E., Nicolaou, K. C., Brunton, L. L., Xuong, N. H., Ten Eyck, L. F. and Taylor, S. S. (1999) Crystal structure of the potent natural product inhibitor balanol in complex with the catalytic subunit of cAMP-dependent protein kinase. *Biochemistry* **38**, 2367–2376
- 111 Kobayashi, E., Nakano, H., Morimoto, M. and Tamaoki, T. (1989) Calphostin C (UCN-1028C), a novel microbial compound, is a highly potent and specific inhibitor of protein kinase C. *Biochem. Biophys. Res. Commun.* **159**, 548–553
- 112 Bruns, R. F., Miller, F. D., Merriman, R. L., Howbert, J. J., Heath, W. F., Kobayashi, E., Takahashi, I., Tamaoki, T. and Nakano, H. (1991) Inhibition of protein kinase C by calphostin C is light-dependent. *Biochem. Biophys. Res. Commun.* **176**, 288–293
- 113 Biondi, R. M., Kieloch, A., Currie, R. A., Deak, M. and Alessi, D. R. (2001) The PIF-binding pocket in PDK1 is essential for activation of S6K and SGK, but not PKB. *EMBO J.* **20**, 4380–4390
- 114 Lopez-Garcia, L. A., Schulze, J. O., Fröhner, W., Zhang, H., Süß, E., Weber, N., Navratil, J., Amon, S., Hindie, V., Zeuzem, S. et al. (2011) Allosteric regulation of protein kinase PKC $\zeta$  by the N-terminal C1 domain and small compounds to the PIF-pocket. *Chem. Biol.* **18**, 1463–1473
- 115 Balendran, A., Hare, G. R., Kieloch, A., Williams, M. R. and Alessi, D. R. (2000) Further evidence that 3-phosphoinositide-dependent protein kinase-1 (PDK1) is required for the stability and phosphorylation of protein kinase C (PKC) isoforms. *FEBS Lett.* **484**, 217–223
- 116 Gould, C. M., Kannan, N., Taylor, S. S. and Newton, A. C. (2009) The chaperones Hsp90 and Cdc37 mediate the maturation and stabilization of protein kinase C through a conserved PXXP motif in the C-terminal tail. *J. Biol. Chem.* **284**, 4921–4935
- 117 Guertin, D. A., Stevens, D. M., Thoreen, C. C., Burds, A. A., Kalaany, N. Y., Moffat, J., Brown, M., Fitzgerald, K. J. and Sabatini, D. M. (2006) Ablation in mice of the mTORC components raptor, rictor, or mLST8 reveals that mTORC2 is required for signaling to Akt-FOXO and PKC $\alpha$ , but not S6K1. *Dev. Cell* **11**, 859–871
- 118 Facchinetti, V., Ouyang, W., Wei, H., Soto, N., Lazorchak, A., Gould, C., Lowry, C., Newton, A. C., Mao, Y., Miao, R. Q. et al. (2008) The mammalian target of rapamycin complex 2 controls folding and stability of Akt and protein kinase C. *EMBO J.* **27**, 1932–1943
- 119 Jacinto, E. and Lorberg, A. (2008) TOR regulation of AGC kinases in yeast and mammals. *Biochem. J.* **410**, 19–37
- 120 Guertin, D. A. and Sabatini, D. M. (2009) The pharmacology of mTOR inhibition. *Sci. Signaling* **2**, pe24
- 121 Abrahamsen, H., O'Neill, A. K., Kannan, N., Kruse, N., Taylor, S. S., Jennings, P. A. and Newton, A. C. (2012) Peptidyl-prolyl isomerase Pin1 controls down-regulation of conventional protein kinase C isozymes. *J. Biol. Chem.* **287**, 13262–13278
- 122 Gao, T., Brognard, J. and Newton, A. C. (2008) The phosphatase PHLPP controls the cellular levels of protein kinase C. *J. Biol. Chem.* **283**, 6300–6311
- 123 Mochly-Rosen, D. and Gordon, A. S. (1998) Anchoring proteins for protein kinase C: a means for isozyme selectivity. *FASEB J.* **12**, 35–42
- 124 Mochly-Rosen, D., Khaner, H. and Lopez, J. (1991) Identification of intracellular receptor proteins for activated protein kinase C. *Proc. Natl. Acad. Sci. U.S.A.* **88**, 3997–4000
- 125 Ron, D. and Mochly-Rosen, D. (1995) An autoregulatory region in protein kinase C: the pseudoanchoring site. *Proc. Natl. Acad. Sci. U.S.A.* **92**, 492–496
- 126 House, C. and Kemp, B. E. (1987) Protein kinase C contains a pseudosubstrate prototope in its regulatory domain. *Science* **238**, 1726–1728
- 127 Churchill, E. N., Qvit, N. and Mochly-Rosen, D. (2009) Rationally designed peptide regulators of protein kinase C. *Trends Endocrinol. Metab.* **20**, 25–33

- 128 Palaniyandi, S. S., Sun, L., Ferreira, J. C. and Mochly-Rosen, D. (2009) Protein kinase C in heart failure: a therapeutic target? *Cardiovasc. Res.* **82**, 229–239
- 129 Shema, R., Sacktor, T. C. and Dudai, Y. (2007) Rapid erasure of long-term memory associations in the cortex by an inhibitor of PKM $\zeta$ . *Science* **317**, 951–953
- 130 Serrano, P., Friedman, E. L., Kenney, J., Taubenfeld, S. M., Zimmerman, J. M., Hanna, J., Alberini, C., Kelley, A. E., Maren, S., Rudy, J. W. et al. (2008) PKM $\zeta$  maintains spatial, instrumental, and classically conditioned long-term memories. *PLoS Biol.* **6**, 2698–2706
- 131 Miguez, P. V., Hardt, O., Wu, D. C., Gamache, K., Sacktor, T. C., Wang, Y. T. and Nader, K. (2010) PKM $\zeta$  maintains memories by regulating GluR2-dependent AMPA receptor trafficking. *Nat. Neurosci.* **13**, 630–634
- 132 Volk, L. J., Bachman, J. L., Johnson, R., Yu, Y. and Huganir, R. L. (2013) PKM- $\zeta$  is not required for hippocampal synaptic plasticity, learning and memory. *Nature* **493**, 420–423
- 133 Lee, A. M., Kanter, B. R., Wang, D., Lim, J. P., Zou, M. E., Qiu, C., McMahon, T., Dadgar, J., Fischbach-Weiss, S. C. and Messing, R. O. (2013) Prkcz null mice show normal learning and memory. *Nature* **493**, 416–419
- 134 Scott, J. D., Glaccum, M. B., Fischer, E. H. and Krebs, E. G. (1986) Primary-structure requirements for inhibition by the heat-stable inhibitor of the cAMP-dependent protein kinase. *Proc. Natl. Acad. Sci. U.S.A.* **83**, 1613–1616
- 135 Alto, N. M., Soderling, S. H., Hoshi, N., Langeberg, L. K., Fayos, R., Jennings, P. A. and Scott, J. D. (2003) Bioinformatic design of A-kinase anchoring protein-*in silico*: a potent and selective peptide antagonist of type II protein kinase A anchoring. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 4445–4450
- 136 Smith, M. K., Colbran, R. J. and Soderling, T. R. (1990) Specificities of autoinhibitory domain peptides for four protein kinases. Implications for intact cell studies of protein kinase function. *J. Biol. Chem.* **265**, 1837–1840
- 137 Nishikawa, K., Toker, A., Johannes, F. J., Songyang, Z. and Cantley, L. C. (1997) Determination of the specific substrate sequence motifs of protein kinase C isozymes. *J. Biol. Chem.* **272**, 952–960
- 138 Gschwendt, M., Muller, H. J., Kielbassa, K., Zang, R., Kittstein, W., Rincke, G. and Marks, F. (1994) Rottlerin, a novel protein kinase inhibitor. *Biochem. Biophys. Res. Commun.* **199**, 93–98
- 139 Soltoff, S. P. (2001) Rottlerin is a mitochondrial uncoupler that decreases cellular ATP levels and indirectly blocks protein kinase C $\delta$  tyrosine phosphorylation. *J. Biol. Chem.* **276**, 37986–37992
- 140 Soltoff, S. P. (2007) Rottlerin: an inappropriate and ineffective inhibitor of PKC $\delta$ . *Trends Pharmacol. Sci.* **28**, 453–458
- 141 Leitges, M., Elis, W., Gimborn, K. and Huber, M. (2001) Rottlerin-independent attenuation of pervanadate-induced tyrosine phosphorylation events by protein kinase C- $\delta$  in hemopoietic cells. *Lab. Invest.* **81**, 1087–1095
- 142 Pula, G., Schuh, K., Nakayama, K., Nakayama, K. I., Walter, U. and Poole, A. W. (2006) PKC $\delta$  regulates collagen-induced platelet aggregation through inhibition of VASP-mediated filopodia formation. *Blood* **108**, 4035–4044
- 143 Shoenfelt, J. L. and Fenton, M. J. (2006) TLR2- and TLR4-dependent activation of STAT1 serine phosphorylation in murine macrophages is protein kinase C- $\delta$ -independent. *J. Endotoxin Res.* **12**, 231–240
- 144 Kurosu, T., Tsuji, K., Kida, A., Koyama, T., Yamamoto, M. and Miura, O. (2007) Rottlerin synergistically enhances imatinib-induced apoptosis of BCR/ABL-expressing cells through its mitochondrial uncoupling effect independent of protein kinase C- $\delta$ . *Oncogene* **26**, 2975–2987
- 145 Herbert, J. M., Augereau, J. M., Gleye, J. and Maffrand, J. P. (1990) Chelerythrine is a potent and specific inhibitor of protein kinase C. *Biochem. Biophys. Res. Commun.* **172**, 993–999
- 146 Lee, S. K., Qing, W. G., Mar, W., Luyengi, L., Mehta, R. G., Kawanishi, K., Fong, H. H., Beecher, C. W., Kinghorn, A. D. and Pezzuto, J. M. (1998) Angoline and chelerythrine, benzophenanthridine alkaloids that do not inhibit protein kinase C. *J. Biol. Chem.* **273**, 19829–19833
- 147 Thompson, L. J. and Fields, A. P. (1996)  $\beta$ II protein kinase C is required for the G2/M phase transition of cell cycle. *J. Biol. Chem.* **271**, 15045–15053
- 148 Yu, R., Mandlekar, S., Tan, T. H. and Kong, A. N. (2000) Activation of p38 and c-Jun N-terminal kinase pathways and induction of apoptosis by chelerythrine do not require inhibition of protein kinase C. *J. Biol. Chem.* **275**, 9612–9619
- 149 Yamamoto, S., Seta, K., Morisco, C., Vatner, S. F. and Sadoshima, J. (2001) Chelerythrine rapidly induces apoptosis through generation of reactive oxygen species in cardiac myocytes. *J. Mol. Cell. Cardiol.* **33**, 1829–1848
- 150 Chan, S. L., Lee, M. C., Tan, K. O., Yang, L. K., Lee, A. S., Flotow, H., Fu, N. Y., Butler, M. S., Soejarto, D. D., Buss, A. D. and Yu, V. C. (2003) Identification of chelerythrine as an inhibitor of BclXL function. *J. Biol. Chem.* **278**, 20453–20456
- 151 Zhang, Y. H., Bhunia, A., Wan, K. F., Lee, M. C., Chan, S. L., Yu, V. C. and Mok, Y. K. (2006) Chelerythrine and sanguinarine dock at distinct sites on BclXL that are not the classic BH3 binding cleft. *J. Mol. Biol.* **364**, 536–549
- 152 Walterova, D., Ulrichova, Preininger, V., Simanek, V., Lenfeld, J. and Lasovsky, J. (1981) Inhibition of liver alanine aminotransferase activity by some benzophenanthridine alkaloids. *J. Med. Chem.* **24**, 1100–1103
- 153 Dvorak, Z. and Simanek, V. (2007) Metabolism of sanguinarine: the facts and the myths. *Curr. Drug Metab.* **8**, 173–176
- 154 Cameron, A. J., Procyk, K. J., Leitges, M. and Parker, P. J. (2008) PKC $\alpha$  protein but not kinase activity is critical for glioma cell proliferation and survival. *Int. J. Cancer* **123**, 769–779
- 155 Shi, F., Telesco, S. E., Liu, Y., Radhakrishnan, R. and Lemmon, M. A. (2010) ErbB3/HER3 intracellular domain is competent to bind ATP and catalyze autophosphorylation. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 7692–7697
- 156 Hu, J., Yu, H., Kornev, A. P., Zhao, J., Filbert, E. L., Taylor, S. S. and Shaw, A. S. (2011) Mutation that blocks ATP binding creates a pseudokinase stabilizing the scaffolding function of kinase suppressor of Ras, CRAF and BRAF. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 6067–6072
- 157 Shah, K., Liu, Y., Deirmengian, C. and Shokat, K. M. (1997) Engineering unnatural nucleotide specificity for Rous sarcoma virus tyrosine kinase to uniquely label its direct substrates. *Proc. Natl. Acad. Sci. U.S.A.* **94**, 3565–3570
- 158 Liu, Y., Shah, K., Yang, F., Witucki, L. and Shokat, K. M. (1998) Engineering Src family protein kinases with unnatural nucleotide specificity. *Chem. Biol.* **5**, 91–101
- 159 Bishop, A. C., Shah, K., Liu, Y., Witucki, L., Kung, C. and Shokat, K. M. (1998) Design of allele-specific inhibitors to probe protein kinase signaling. *Curr. Biol.* **8**, 257–266
- 160 Cohen, P. and Goedert, M. (1998) Engineering protein kinases with distinct nucleotide specificities and inhibitor sensitivities by mutation of a single amino acid. *Chem. Biol.* **5**, R161–R164
- 161 Cibrrian Uhalte, E., Kirchner, M., Hellwig, N., Allen, J. J., Donat, S., Shokat, K. M., Selbach, M. and Abdelilah-Seyfried, S. (2012) *In vivo* conditions to identify Prkci phosphorylation targets using the analog-sensitive kinase method in zebrafish. *PLoS ONE* **7**, e40000
- 162 Irie, N., Sakai, N., Ueyama, T., Kajimoto, T., Shirai, Y. and Saito, N. (2002) Subtype- and species-specific knockdown of PKC using short interfering RNA. *Biochem. Biophys. Res. Commun.* **298**, 738–743
- 163 Justilien, V., Jameison, L., Der, C. J., Rossman, K. L. and Fields, A. P. (2011) Oncogenic activity of Ect2 is regulated through protein kinase C $\alpha$ -mediated phosphorylation. *J. Biol. Chem.* **286**, 8149–8157
- 164 Abeliovich, A., Chen, C., Goda, Y., Silva, A. J., Stevens, C. F. and Tonegawa, S. (1993) Modified hippocampal long-term potentiation in PKC $\gamma$ -mutant mice. *Cell* **75**, 1253–1262
- 165 Leitges, M., Schmedt, C., Guinamard, R., Davoust, J., Schaal, S., Stabel, S. and Tarakhovskiy, A. (1996) Immunodeficiency in protein kinase C $\beta$ -deficient mice. *Science* **273**, 788–791
- 166 Bandyopadhyay, G., Standaert, M. L., Sajjan, M. P., Kanoh, Y., Miura, A., Braun, U., Kruse, F., Leitges, M. and Farese, R. V. (2004) Protein kinase C- $\lambda$  knockout in embryonic stem cells and adipocytes impairs insulin-stimulated glucose transport. *Mol. Endocrinol.* **18**, 373–383
- 167 Hodge, C. W., Mehmert, K. K., Kelley, S. P., McMahon, T., Haywood, A., Olive, M. F., Wang, D., Sanchez-Perez, A. M. and Messing, R. O. (1999) Supersensitivity to allosteric GABA(A) receptor modulators and alcohol in mice lacking PKC $\epsilon$ . *Nat. Neurosci.* **2**, 997–1002
- 168 Choi, D. S. and Messing, R. O. (2003) Animal models in the study of protein kinase C isozymes. *Methods Mol. Biol.* **233**, 455–473
- 169 Jackson, D., Zheng, Y., Lyo, D., Shen, Y., Nakayama, K., Nakayama, K. I., Humphries, M. J., Reyland, M. E. and Foster, D. A. (2005) Suppression of cell migration by protein kinase C $\delta$ . *Oncogene* **24**, 3067–3072
- 170 Shu, X., Lev-Ram, V., Deerinck, T. J., Qi, Y., Ramko, E. B., Davidson, M. W., Jin, Y., Ellisman, M. H. and Tsien, R. Y. (2011) A genetically encoded tag for correlated light and electron microscopy of intact cells, tissues, and organisms. *PLoS Biol.* **9**, e1001041
- 171 Sutton, R. B. and Sprang, S. R. (1998) Structure of the protein kinase C $\beta$  phospholipid-binding C2 domain complexed with Ca<sup>2+</sup>. *Structure* **6**, 1395–1405